# Selenium in Seleniferous Environments

# H. F. Mayland

USDA-Agricultural Research Service Kimberly, Idaho

L. F. James and K. E. Panter

USDA-Agricultural Research Service Logan, Utah

J. L. Sonderegger

Montana Bureau of Mines and Geology Butte, Montana

# ABSTRACT

Selenium is biologically important because (i) it is essential in animal and possibly plant metabolism, (ii) in many areas diets do not contain sufficient Se to meet animals' needs, and (iii) in other areas it is toxic to animals when it occurrs in high concentrations in soil, water, plants, fly ash, or in aerosols. Animals require 0.05 to 0.1 mg Se/kg in their diets to prevent Se deficiency but suffer Se toxicosis when dietary levels exceed 5 to 15 mg Se/kg. The earth's crustal materials generally contain < 0.1 mg Se/kg. Higher concentrations are found in Cretaceous shales. The Seaccumulator plants growing on the seleniferous soils may contain hundreds or even thousands of mg Se/kg. However, the nonaccumulator grasses and forbs seldom accumulate > 50 mg Se/kg and more often contain < 5 mg Se/kg. Soils and plants may discharge volatile forms of Se into the atmosphere. However, plants may also absorb measurable amounts of gaseous Se from the atmosphere. Anthropogenic activities impact the amount of Se entering our nation's lakes, rivers, and the atmosphere. Combustion of coal and incineration of municipal waste exhaust Se into the environment. In addition, crop-fallow and irrigation practices that allow leaching waters to pass through seleniferous strata prior to intersecting with surface flow, augment the Se levels encountered by plant and animal life.

In 1857, U.S. Army Surgeon T.W. Madison described toxicity symptoms in horses (*Equus caballus*) grazing near Fort Randall, SD (Rosenfeld & Beath, 1964). Since then, ranchers in that area have continued to experience livestock losses. They associated the toxicosis with the saline seeps and outcrops, com-

Copyright © 1989 Soil Science Society of America and American Society of Agronomy, 677 S. Segoe Rd., Madison, WI 53711, USA. Selenium in Agriculture and the Environment, SSSA Special Publication no. 23. mon to much of the area, and named the problem *alkali disease*. Not until 1931 did researchers associate alkali disease with selenosis. Since then, researchers have continued to characterize Se in soils and plants and its toxicosis in animals. Early efforts succeeded in describing areas in the western USA and Canada where Se accumulator plants contained concentrations of 50 to 500 mg Se/kg or higher (Rosenfeld & Beath, 1964; Fig. 2-1).

Research continued on soil, plant, and animal factors associated with high Se levels, but by 1958, nutritional deficiencies of Se were noted in the diets of many animals including grazing livestock (NAS-NRC, 1983). Areas of the USA were then mapped where plants contained <0.1 mg Se/kg (Fig. 2–2) and where Se deficiency disorders were likely to occur in livestock and poultry. In general, Se concentrations of 0.05 to 0.1 mg Se/kg diet and 5 to 15 mg Se/kg diet may be considered as critical levels of adequacy and toxicity, respectively.



Fig. 2-1. Distribution of seleniferous vegetation in the western USA and Canada (adapted from Rosenfeld & Beath, 1964). Each open dot represents the place of collection of a plant specimen containing 50-500 mg Se/kg; each solid dot represents specimens containing > 500 mg Se/kg.



Fig. 2-2. Locations and classification of counties where plants were collected for Sc analysis (adapted from Kubota et al., 1967). Vertical lines, X's and solid dots indicate counties where more than one-half of the crops sampled contained <0.1, >0.1, or where Se-accumulator plants (data on forages and grains were unavailable) contained >50 mg Se/kg, respectively.

MAYLAND ET AL.

Knowledge of Se in the environment prior to the mid-1970s has been summarized in a number of excellent reviews: Allaway et al. (1967), Allaway (1968), Anderson et al. (1961), Anderson and Scarf (1983), Brown and Shrift (1982), Gissel-Nielsen et al. (1984), Koller and Exon (1986), Martin (1973), Moxon and Rhian (1943), NAS-NRC (1974, 1976, 1983), Reuter (1975), Sharma and Singh (1983), and Shrift (1958, 1969, 1973). Selenium information is also given in the recent publications by Adriano (1986), Combs and Combs (1986), Ihnat (1989), and Levander (1986).

Since the mid-1970s human activities have introduced new sources of Se into the ecosystem, and much attention is now being given to the anthropogenic impact of Se cycling in seleniferous environments. This chapter will briefly describe Se in the various compartments that were characterized through the mid-1970s but will elaborate in more detail on the impact of human activities. However, those aspects associated with the central California experience will be detailed in other chapters of this special publication.

Low, medium or adequate, and high levels of bioavailable Se, as used in this chapter, will refer to those levels in the soil that result in concentrations of < 0.1, 0.1 to 1.0, and > 1.0 mg Se/kg, respectively, in the non-Seaccumulator plants growing on such soils.

# GEOCHEMISTRY

Selenium occurs in nearly all materials of the earth's crust and is present in magmatic rocks in concentrations generally <0.05 mg/kg (Table 2-1). During volcanic activity, much of the Se escapes as high-temperature volatile gases (Reuter, 1975); thus, igneous rocks are usually low in Se. The element is rarely seen in elemental form (Se<sup>0</sup>) but may be found in association with sulfide ore bodies (Anderson et al., 1961).

Selenium concentrations are higher in sedimentary rocks where it is associated with the clay fraction, and thus, larger concentrations of Se are found in shales than in limestones or sandstone. Even in shales, the concentrations

Rock type	Se conc., mg/kg
Magmatic rocks	
Ultramafic rocks (Dunites, Pendotites, Pyroxeuites)	0.02-0.05
Mafic rocks (Basalts, Gabbros)	0.01-0.05
Intermediate rocks (Diorites, Syenites)	0.02-0.05
Acid rocks (Granites, Gueissas)	0.01-0.05
Acid rocks-volcanic (Rhyolites, Trachytes)	0.02-0.05
Sedimentary rocks	
Argillaceous sediments	0.4-0.6
Shales	0.6
Sandstones	0.05-0.08
Phosphorites	1-100
Limestones, dolomites	0.03 - 0.10

Table 2-1. Selenium concentration in magmatic and sedimentary rocks (compiled from Kabata-Pendias & Pendias, 1984).

are seldom > 500 mg Se/kg. Values range from <1 to nearly 300 mg Se/kg in the Cretaceous black shales in Wyoming (NAS-NRC, 1983). Approximately 2 mg Se/kg occurs in the Cretaceous Pierre and related shales, which are the parent material for much of the seleniferous soil in the northern Great Plains of the USA and the Prairie region of Canada. These seleniferous soils may have higher concentrations of Se in the subsoil because of chemical weathering. Shales are also the principal sources of Se-toxic soil in Ireland, Australia, Israel, and several other countries of the world (Abuereish & Lahham, 1987; Anderson et al., 1961; Lakin, 1948; Rosenfeld & Beath, 1964; Swaine, 1955; NAS-NRC, 1983).

The relatively high concentration of Se in some phosphate rocks (Rader & Hill, 1935), particularly western phosphates (1-178 mg Se/kg), is significant because of the wide use of phosphate fertilizers. Robbins and Carter (1970) suggested that normal superphosphate can be expected to retain about 60% and concentrated superphosphate about 40% of the Se found in the original rock. The decrease in the Se concentration results from volatilization and loss of some Se during the smelting process.

Coal often contains from 1 to 20 mg Se/kg, although some coal from the People's Republic of China has >1000 mg Se/kg (O.E. Olson, 1986, personal communication). Upon combustion, coal releases large amounts of Se into the ecosystem. This topic will be covered in more detail later in this chapter. Crude oil also contains Se. Ellrich et al. (1985) measured the Se and S concentrations in 45 crude oils of southern West Germany and reported a range of 0.02 to 0.2 mg Se/kg. The Se and S values were highly correlated.

There are no known ore deposits sufficiently high in Se to permit mining it exclusively. Most commercially available Se is extracted from copper refinery slime along with the recovery of other precious metals (NAS-NRC, 1976, 1983). Total production of Se in the world was  $1.89 \times 10^6$  kg Se in 1983, including  $200 \times 10^3$  kg in the USA (Jensen, 1985). Production by continent was as follows:

North America	$645 \times 10^3 \text{ k}$	g
South America	330	
Europe	395	
Africa	230	
Asia	190	
Oceana	95	

Selenium is used in the manufacture of batteries, glass, and electronic equipment, and as a reagent in remedies for eczema, fungus infections, antidandruff products, veterinary therapeutic agents, injectables, feed additives, and fertilizers. The demand for fertilizer Se may exceed  $28 \times 10^3$  kg by 1990 (Korkman, 1985).

# SELENIUM IN SOIL

The concentration of Se in most soils lies within the range of 0.01 to 2 mg Se/kg. However, high concentrations of up to 1200 mg Se/kg total

and 38 mg Se/kg as water-soluble selenate have been reported in seleniferous areas of the world (Lakin, 1948; Swaine, 1955). Soils developed from the Cretaceous shales of South Dakota, Montana, Wyoming, Nebraska, Kansas, Utah, Colorado, and New Mexico tend to have high Se values ranging from 1 to >10 mg Se/kg. It is in these areas where natural Se toxicosis has occurred historically. The Se content of some Hawaiian surface soils varies from 1 to 20 mg/kg, but this Se is unavailable for vegetation because of it complexation with the Fe and Al minerals (Anderson et al., 1961; Rosenfeld & Beath, 1964).

Low-Se soils in the USA include those derived from sedimentary rocks that predate the major Cretaceous period (northeastern USA). Other low-Se soils include those that are formed from recent volcanic ash deposits or loessal material derived from the ash (Pacific Northwest). Soils in the very low-Se region of the eastern USA are formed from coastal deposits consisting of weathered, recycled sediments (South Atlantic seaboard).

The soil parent materials of the low-Se areas in Montana and parts of Idaho are mostly derived from granites and old metamorphic rocks. Low total-Se concentrations occur in the tertiary volcanic rocks of Arizona and New Mexico. Most of the soils from low-Se areas of the USA contain <0.5 mg Se/kg. Many of these low-Se areas correspond to areas where Sedeficiency disorders such as white muscle disease occur.

As with other elements, the total concentration of Se in soils shows little relationship to the concentration of Se in plants grown on those soils (Lakin, 1972). This is because Se in soil exists in several chemical forms that differ widely in their solubility and availability to plants (Anderson & Scarf, 1983). These chemical forms are:

Selenide,  $Se^{2-}$ ,  $HSe^{-}$ ,  $H_2Se$  (aq) Elemental selenium,  $Se^{0}$ Selenite,  $SeO_3^{2-}$ ,  $HSeO_3^{-}$ ,  $H_2SeO_3$  (aq) Selenate,  $SeO_4^{2-}$ ,  $HSeO_4^{-}$ ,  $H_2SeO_4$  (aq) Organic Se

The chemical forms of Se present in soils and sediments are closely related to the oxidation-reduction potential and pH of the soil (van Dorst & Peterson, 1984, Fig. 2-3). Inorganic Se exists predominately as the oxyanions, selenite and selenate, in the aqueous solutions of well-aerated alkaline soils. In poorly aerated acid soils, inorganic Se predominates as the relatively insoluble selenide and elemental forms. Howard (1977) summarized the Se geochemistry, as affected by Fe, and concluded that the geochemistry of Se is largely controlled by that of Fe, with which Se is closely affiliated in both oxidizing and reducing environments.

Soils will be high in biologically available Se if formed from parent materials that were high in Se and maintained under alkaline conditions. Plants growing on these soils will have sufficient Se (0.1 mg Se/kg forage) for animal requirements. However, if the bioavailability of the Se is very high, it may produce Se toxicosis in plants or the animals that ingest them (Smith & Watkinson, 1984). Soltanpour and Workman (1980) indirectly ex-



Fig. 2-3. Variation of oxidation potentials for the predominating states of Se as a function of pH. A line intersecting the point Eh = 0.4 and pH = 4 and the point Eh = 0.1 and pH = 9 divides the oxidized soils above and the reduced soils below (adapted from Lakin & Davidson, 1967 and NAS-NRC, 1976).

amined the bioavailability of soil Se by using ammonium bicarbonate-DTPA (diethylenetriaminepenta acetic acid) as a soil extractant. Alfalfa (*Medicago sativa* L.) contained in excess of 5 mg Se/kg when growing on mollisols containing more than 45 mg extractable Se/kg soil. Alfalfa yields were reduced when the extractable soil Se exceeded 500 mg/kg. The acutal amount of Se for toxicosis will depend on species of plant, species and age of animal, and other factors including concentrations of SO<sub>4</sub>-S, P, and sesquioxides in the soil.

Soils that were formed from either high or low Se parent materials, but have poor aeration; acid pH; and high rainfall are likely to be soils that may be deficient in either total or at least bioavailable Se. Plants growing on these latter soils may not contain sufficient Se to meet animal requirements. More detail on the uptake and accumulation of Se by agricultural crops is presented in Chapter 4 of this publication.

## SELENIUM IN WATER

Selenium occurs as a minor constituent in drinking water in a concentration range of <0.1 to 100  $\mu$ g/L (NAS-NRC, 1983). Samples rarely exceed the 10  $\mu$ g/L upper limit established by the 1977 Safe Drinking Water Act of the USEPA. However, water derived from some of the Cretaceous zones may have high concentrations of both Se and total soluble salts. For example, water from wells drilled into any of the geologic formations of the Cretaceous Colorado group in central Montana may contain as much as 1000  $\mu$ g Se/L (Donovan et al., 1981). The total dissolved solids in this water average 9260 mg/L.

Rumble (1985) measured water quality parameters, including Se, in surface-coal and bentonite-mine impoundments and livestock ponds in the Northern High Plains. He reported mean Se values of 2.2, 10.6, and 1.1  $\mu$ g Se/L in the three potential livestock-water sources, respectively. H.F. May-land (1986, unpublished data) also sampled livestock-water impoundments throughout central Montana in known seleniferous areas and found 0.4 to 0.7  $\mu$ g Se/L in most of these waters. A few samples contained as high as 270  $\mu$ g Se/L, many times greater than the USEPA (1977) allowable value of 10  $\mu$ g Se/L in drinking water but is near the 100  $\mu$ g/L level of Se in blood considered adequate for animal requirements.

In a report on water-quality trends in the nation's rivers, Smith et al. (1987) noted that, over an 8-yr period, 75% of the 211 stations reported values at or below the 1  $\mu$ g Se/L laboratory detection limit. Four sites had higher concentrations in 1981 than in 1974, whereas 23 had lower concentrations. Rivers draining some of the seleniferous regions contained relatively high Se concentrations. Selenium mean values and trends for specific stations (R.A. Smith, 1987, personal communication, Table 2-2) show a decline in flow-corrected values of Se in three rivers flowing out of seleniferous areas. These decreases in Se outflow were also accompanied with decreases in total dissolved salts (TDS) from the two Colorado sites. The TDS information was not available for the Montana site. Improvements in irrigation water management in western Colorado and modifications of the crop-fallow system in north-central Montana have occurred during this same period. These practices would account for decreases in river-Se levels if also accompanied by decreases in overall salinity.

Smith et al. (1987) reported an increasing trend in arsenic (As) and cadmium (Cd) concentrations in many of the rivers. Fossil-fuel combustion, the largest source of both elements, introduces these elements into the aquatic environment through (i) atmospheric deposition of combustion products and (ii) runoff from fly ash storage areas near power plants and other nonferrous smelters. Fossil-fuel combustion also releases Se to the environment

cation,					
		Dissolved Se		Total Se	
River station	Location	Conc.	Trend†	Conc.	Trend†
		$\mu { m g}/{ m L}$		$\mu g/L$	
Gunnison River	Near Grand Junction, CO	8.0	Down	10.0	Down
Colorado River	Colorado-Utah boundary	5.5	Down	8.5	Down
Marias River	Chester, MT	1.0	$\mathbf{Down}$	1.0	Down
Chevenne River	Cherry Creek, SD	3.0	NS	4.0	NS
Little Missouri	Waterford City, ND	1.0	NS	1.0	NS
Missouri River	Fort Randall, SD	2.0	NS	2.0	NS

Table 2-2. Mean Se values and concentration trends over the 1974 through 1981 sampling period for river systems in seleniferous areas (R.A. Smith, 1987, personal communication).

† Trends are significant at P < 0.10. NS = not significant.

where it could eventually appear as fallout. However, Se concentrations in many rivers may be less than the present detection limits, so changes as noted for As and Cd may not yet be measurable.

# **SELENIUM IN PLANTS**

### Accumulators versus Nonaccumulators

Rosenfeld and Beath (1964) and Shrift (1973) divided plants into three groups on the basis of their ability to accumulate Se when grown on high-Se soils. The first two groups of plants are referred to as *Se accumulator* or *indicator plants*. These grow well on soil containing high levels of available Se, and some have been used to locate seleniferous soils. Plants in Group 1 are called *primary indicators* and include many species of *Astragalus*, *Machaeranthera*, *Haplopappus*, and *Stanleya*. These species absorb high concentrations of Se that may be in the hundered or occasionally even thousands of milligrams per kilogram, dry wt. (Fig. 2-4).

Plants in Group 2 are referred to as secondary Se absorbers. They belong to a number of genera including Aster, some species of Astragalus, Atriplex, Castilleja, Grindelia, Gutierrezia, some species of Machaeranthera, and Mentzelia. They rarely concentrate more than 50 to 100 mg Se/kg. Plants in Group 3 include grains, grasses and many forbs that do not usually accumulate Se in excess of 50 mg Se/kg when grown on seleniforous soil.



Fig. 2-4. Proportion of samples having less than given Se concentration (%). (A) ▲ - ▲ Data for western wheatgrass (*Pascopyrum smithii* [Rydb.] A. Love) and Sandberg bluegrass (*Poa secunda* Presl.) sampled from seleniferous areas of Montana and Wyoming by H.F. Mayland, (1987, unpublished data); △ - △ Stanleya spp.; ● - △ Xylorhiza section of Machaeranthera; □ - □ Astragalus bisulcatus; X--X Astragalus pectinatus; and ○ - ○ Oonopsis section of Haplopappus; latter data adapted from Rosenfeld & Beath (1964). (B) ● - ● Data are for vegetative wheat (*Triticum aestivum* L.); ■ - ■ Astragalus bisulcatus; and △ - △ Astragalus pectinatus; and △ - △ Astragalus pectinatus;

Some plants growing on seleniferous soils accumulate surprisingly low levels of Se. White clover (*Trifolium repens* L.), buffalograss (*Buchloe dactyloides* [Nutt.] Engelm.), and grama (*Bouteloua* sp.) are poor accumulators of Se. On the other hand, high sulfur (S)-containing plants like the *Brassica* sp. (mustard, cabbage, broccoli, and cauliflower) and other Cruciferae are relatively good concentrators of Se (NAS-NRC, 1983). Absorption of Se and S by plants may be correlated (Shrift, 1973).

# Plant Response to Selenium Species

Gissel-Nielsen and Bisbjerg (1970) evaluated Se uptake by agricultural plants from various Se compounds (Table 2-3). They found in greenhouse and field studies that over a 2-yr period, red clover (*Trifolium pratense* L.), barley (*Hordeum vulgare* L.), and white mustard (*Sinapis alba* L.) absorbed <0.1% of the applied elemental Se, about 1.0% of the SeO<sub>2</sub> and Selepite forms, and 12 to 60% of the Se added as selenate (Table 2-3). In another study, Se uptake by alfalfa, barley, and sugar beet (*Beta vulgaris* L.) was one-third or less of that taken up by the *Cruciferous* mustard. Uptake by red clover decreased by a factor of six over the eight successive harvests regardless of the Se source. Selenium concentrations in clover grown on the selenate sources were 25 to 50 times that obtained from selenite, and the decrease in selenate uptake with time was about four times greater.

In another study, Gissel-Nielsen (1973) again reported that more Se was taken up from selenate than selenite by barley and red clover. With subsequent cropping, there was a greater decline in uptake from the <sup>75</sup>Se labeled sodium selenate source than from the labeled sodium selenite source. Adding sulfate ( $SO_4^{2-}$ ), at rates of 0 to 375 mg S/kg soil, greatly decreased the uptake of Se from the selenate source but had a lesser effect on Se uptake from the selenite source. The ratio of Se in roots to Se in tops shows that Se is more readily translocated from the roots when taken up from added selenate than from added selenate.

	Calubility of Co	So added	Uptake relative to added Se		
Se source	in cold water	to soil	Clover	Barley	Mustard
	g/L	mg/kg		%	<u> </u>
Se	i	2.5	0.005	0.02	0.07
SeO <sub>2</sub>	i	0.5	1.0	0.9	1.2
K <sub>o</sub> SeO <sub>2</sub>	22.4	0.5	1.0	1.1	1.3
Na <sub>2</sub> SeÖ <sub>3</sub>	S	0.5	1.0	1.0	1.1
BaSeO <sub>2</sub>	0.05	0.37	0.9	0.9	0.9
FeSeO,	i	0.35	1.1	1.0	1.1
CuSeO3	i	0.30	0.8	0.8	0.7
K <sub>o</sub> SeO <sub>4</sub>	390	0.50	24.	12.	24.
BaSeO	0.03	0.10	63.	27.	61.
CuSeO <sub>4</sub>	68	0.13	53.	28.	48.

Table 2-3. Selenium solubility in water and relative uptake of Se by plants from different sources labeled with <sup>75</sup>Se in pot experiments using a loamysand having 2.8% organic matter, 5.7 pH, and 0.12 mg Se/kg (adapted from Gissel-Nielsen & Bisbjerg, 1970).

 $\dagger i = insoluble, s = slightly soluble.$ 

Hamilton and Beath (1964) grew vegetable plants in soil-pots containing either 3 mg Se/kg as selenate or 20 mg Se/kg as finely ground powder of the Se accumulating plants, *Astragalus bisulcatus* or *A. preussii*. Vegetable plants grew satisfactorily and most accumulated higher concentrations of Se from the organic Se source, probably because a greater amount of bioavailable Se was produced by the larger application. The absorbed Se was stored in soluble organic and inorganic forms. Selenate, but not selenite, was detected in these test plants.

Selenite, selenate, and organic Se at several rates were included in another study (Hamilton & Beath, 1963) of Se uptake involving rangeland forbs and grasses. Selenium from the selenate form was generally absorbed more efficiently than that from the selenite form. Selenium from the seleniferous plant source was absorbed least efficiently. However, plants identified in Groups 1 and 2 (see above) absorbed comparatively large amounts of organic Se from the soil.

Plant uptake of selenate-Se occurs at higher rates than that of the selenite-Se. Peterson et al. (1981) suggested that selenate may be taken up actively by plants, whereas selenite may be absorbed passively and in lesser concentrations. Nevertheless, it is the selenate source that is subject to competition from  $SO_4^{2-}$  and other anions.

# Plant Response to Low Bioavailable Selenium

Of plants growing on moderately low-Se soil, alfalfa accumulates more Se than does red clover, timothy (*Phleum pratense* L.) or brome grass (*Bromus inermis* Leyss.) (Massey & Martin, 1975). Differences in Se uptake may occur, because some plants like alfalfa are deeply rooted and may access more bioavailable Se in the deeper, more alkaline subsoil horizons. No definite differences in Se accumulations have been noted among species growing on very low levels of available Se.

Crops growing on neutral or acid soil absorb very little Se, and any attempt to increase crop Se uptake by shifting to some other species has been viewed with some skepticism (Ehlig et al., 1968). However, small but significant differences have been noted among selection of tall fescue growing on soil having moderately low levels of available soil Se (preliminary work by D. Sleper and H.F. Mayland, 1987). Therefore, some opportunity may exist to breed forage grasses that would take up greater concentrations of Se and could be planted in areas now considered to be Se-deficient for animals.

Davies and Watkinson (1966) reported that a marked difference in Se concentration between species was apparent under New Zealand conditions of low-Se soil. The Se concentration was greatest in a native grass (Agrostis tenuis Sibth.) and least in white clover (Trifolium repens L.), reflecting possible genetic control of Se absorption and transport to the plant tops. The Se values for orchardgrass (Dactylis glomerta L.) and ryegrass (Lolium perenne L.) fell between these extremes.

The Se in forages and cereal grains may not differ much among species in areas where available Se levels in the soil are low. Selenium concentrations in legumes and grasses vary when these plants are grown on soils where bioavailability of Se to forages is adequate for animal requirements. However, alfalfa usually contains the highest level, which may be several times as much as is in the grass forages (Massey & Martin, 1975). In these same areas, Se levels in wheat (*Triticum aestivum* L.) grain have been shown to be higher than in oat (*Avena sativa* L.) or barley.

Westermann and Robbins (1974) noted that, in many areas where S is deficient for plant growth, Se may also be inadequate for animal requirements. Application of fertilizer S to increase forage yields may further lower forage Se concentrations either by direct competition between  $SO_4^{2-}$  and  $SeO_4^{2-}$  at the soil root interface, by stimulation of growth and dilution of Se, or both.

Williams and Thornton (1972) evaluated, in the glass house, the effect of several soil additives on the plant uptake of Se from soils containing <4to 230 mg Se/kg. Ammonium sulphate and elemental S markedly decreased the Se uptake by perennial ryegrass and red clover, with the ammonium salt giving the most persistent effect. Both the ammonium and the sulphate ions seem to be involved. Calcium orthophosphate and potassium sulphate sometimes increased Se uptake, so use of these fertilizers on Se-toxic soils should be considered with caution.

## Plant Response to High Bioavailable Selenium

Selenium is known to be required for animal health (0.05-0.1 mg Se/kg diet) but is not yet considered as an essential element for plant growth. Several earlier studies strongly suggested that Se was required at least for the accumulator plant species. Trelease and Trelease (1938, 1939) reported a pronounced stimulating effect in the growth of Astragalus racemosus (Pursh) and A. pattersonii (Gray) ex. Brand, grown in nutrient cultures containing up to 9 mg Se/L as selenite. Both species are identified as Se accumulator plants. Astragalus crassicarpus Nutt, (also named A. succulentus by Rosenfeld & Beath, 1964), which is a nonaccumulator, was poisoned by Se at rates as low as 0.3 mg/L. These findings led the Treleases' and others (Rosenfeld & Beath, 1964; Shrift, 1969) to suggest that Se may be an essential element for the Se indicator plants. This apparent Se requirement by the accumulator species may be confounded by a Se  $\times$  P interation, whereby the increased growth in the presence of Se could have been due to a depression of P toxicity by selenite (Brover et al., 1972a). However, evidence for the  $P \times Se$  interaction was obtained using solution culture, and proof would be difficult to establish using soil cultures.

In an in-depth, solution-culture study of ionic sorption interactions in *Astragalus* sp., Broyer et al. (1972b) reported the following observations:

... selenite tended to depress the sorption of phosphate and the micronutrient metals (Cu, Fe, Mn, Zn); phosphate tended to restrict selenite sorption and phosphate tended to enhance the sorption of the macro-

nutrient cations and the micronutrient metals in interactions which often resulted in yield differences. Where the influences were relatively well balanced, plant growth was favorable and yields were relatively constant.

Broyer et al. (1972a) observed floral initiation in 6 of 25 replicate Astragalus bisulcatus grown in 20  $\mu$ g Se/L as selenite, whereas no flowering occurred in plants grown in the control cultures. This result, indicating a possible beneficial effect of Se, was also noted by Trelease and Trelease (1939) with A. racemosus. However, Broyer et al. (1972a) believed that the relatively large growth benefit from Se application reported by them for A. racemosus was at least in part, possibly related, to corrected phosphate toxicity in solution culture.

If either A. bisulcatus (accumulator) or A. crotolariae (nonaccumulator) has a requirement for Se, the critical level probably would be  $< 80 \ \mu g$ Se/kg dry plant tops (Broyer et al., 1972a). This value is of the same order reported for alfalfa and/or subterranean clover (*Trifolium subterraneum* L.) (Broyer et al., 1966, 1972a). These values are much less than the comparable data suggested by Trelease and Trelease (1939) and Rosenfeld and Beath (1964), which were 3000 times and 15 000 times greater for the two legume crops, respectively.

# Selenium Compounds in Plants

Shift (1973) has summarized the findings on the many chemical compounds of Se isolated from plants. Much of the Se in nonaccumulating species is found in the form of protein-bound selenomethionine. In contrast, the Se in accumulator plants is mostly water-soluble and found in nonprotein forms like Se-methylselenocysteine. Only trace amounts of the methylselenocysteine are found in the nonaccumulator species. Metabolic pathways of Se in plants probably diverge at the Se-cysteine stage to go toward proteinbound selenomethionine in nonaccumulators and to the nonprotein forms like methylselenocysteine in accumulator plants (Shrift, 1969, 1973).

Lewis (1976) summarized the metabolic differences between Se accumulators and nonaccumulators. Selenomethionine, selenocystine, and possibly Se-methyl selenomethionine and selenonium have been detected in nonaccumulators but not in the accumulators that have been tested. Accumulator species can form selenocystathionine from selenate or selenite. The subsequent formation of methylselenocysteine has been found to occur only in accumulators and has been suggested as a biochemical basis for distinguishing accumulators from nonaccumulators.

Although little incorporation of the selenoamino acids into proteins occurred in the accumulator plants tested (Peterson & Butler, 1962), extensive protein incorporation of selenoamino acids occurred in ryegrass, wheat, and clover. Lewis (1976) suggested that Se accumulator species have evolved a detoxification mechanism whereby Se is excluded from protein incorporation, whereas nonaccumulator species do not have this mechanism. Selenium incorporated into proteins could result in alteration of the protein structure, inactivation of the protein, and eventual poisoning of the plant (Peterson & Butler, 1962). Selenium-containing transfer ribonucleic acids (tRNA) have been found in a number of aerobic and anaerobic bacteria, mammalian cells, and in Seaccumulating and nonaccumulating plants (Wen et al., 1988). The Secontaining tRNA appears to be ubiquitous in the plant kingdom. However, the distribution of selenonucleoside(s) in tRNA species varies both qualitatively and quantitatively in different plants. This information provides evidence for a possible biological role of Se in higher plants. Nevertheless, the possibility that Se substituted for S in the nucleoside has not been evaluated.

The Se metabolites in plants are, generally, analogs of S compounds. Nevertheless, Se metabolism in non-Se-accumulator plants cannot be identified from known mechanisms involving Se metabolism (Shrift, 1973). Our understanding of the metabolic pathways for Se in plants remains very limited.

Many microorganisms can reduce selenite to elemental Se. Some bacteria and fungi have been found that reduce selenite or selenate to volatile, organic Se compounds (Shrift, 1973). Sulfur-Se antagonism occurs in these microorganisms. The reduction of selenite is enzymatically mediated, and many isolated enzyme systems can utilize S and Se analogs interchangeably. However, what was traditionally thought to be the mechanism of Se toxicity, namely, a general interference with the enzymes involving S assimilation, has proved to be more complex.

# Selenium Toxicity in Plants

The toxicity of Se on the growth of grasses was noted as early as 1880 (Brown & Shrift, 1982). However, the concern about selenosis in plants did not really develop until the identification of selenosis in livestock and the probable Se-associated reductions in wheat production in the seleniferous areas (O.E. Olson, 1986 personal communication).

Selenium toxicity to nonaccumulator plants was described by Singh and Singh (1978). They treated soil in the greenhouse with 0, 2.5, 5, or 10 mg Se/kg as selenite and measured relative wheat-grain production of 100, 72, 17, and 11%, respectively. The grain contained 0.6, 10.2, 24.5, and 36.8 mg Se/kg, respectively. The application of Se to the soil decreased the growth of wheat and sunflower (*Helianthus annuus* L.); the absorption of Zn, Cu, Fe, Mn, S, and N; and the synthesis of nucleic acids, chlorophyll, and protein, whereas additions of 50 or 100 mg P/kg as  $K_2PO_4$  decreased the harmful effect of Se and increased the values of each of the parameters tested.

Other plants, namely those classified by Rosenfeld and Beath (1964) as primary or secondary indicator plants, tolerate and possibly require Se for growth. These indicator plants differ from the nonaccumulator species in several ways. Large quantities of Se-methylselenocysteine and selenocystathionine, two nonprotein selenoamino acids rarely detected in nonaccumulators, have been isolated from the tissues of Se accumulators (Brown & Shrift, 1982). Also, Se is kept from entering proteins so that the Se level in proteins of accumulator plants is significantly lower than the level in Se-sensitive plants. Exclusion of Se from the proteins of accumulators is thought to be the basis

of Se tolerance. Selenium absorbed by the nonaccumulator plants is converted into Se metabolites, which act as analogues of essential S compounds and interfere with cellular biochemical reactions.

# SELENIUM IN ANIMALS

Selenium toxicosis in aquatic and terrestrial animals has been well described in several reviews (NAS-NRC, 1983; Sorensen, 1986; Chapter 8 of this publication). Selenite, selenate, and organic Se sources produce similar toxicological effects (Moxon & Rhian, 1943; unpublished data by L.F. James et al., 1986). Absorption rates vary between chemical forms. Minimum lethal doses may be 1.5 to 3.0 mg Se/kg body weight as selenate (Moxon & Rhian, 1943), although tolerance levels differ between and within animal species and nutritional conditions (Volcani et al., 1957).

Chronic toxicity studies have indicated that diets providing 4 to 5 mg Se/kg body weight or more of Se daily result in chronic toxicity in laboratory animals (NAS-NRC, 1983). The Food and Nutrition Board of the National Academy of Sciences (1980) has accepted 5 mg Se/kg diet as the critical level between toxic and nontoxic feeds.

In general, Se as selenomethionine is more readily absorbed when ingested by animals than is selenite, selenate, or selenocystine (NAS-NRC, 1983). Nearly one-half of the Se in wheat grain is present as selenomethionine (Olson et al., 1970), so the Se in wheat is more readily available to animals than is the Se in selenocystine, *Astragalus*, or fish meal. Generally, Se from plant forms is more available to animals than Se from animal forms.

As in plants, Se in animals interacts with other trace elements. Frank et al. (1986) found 20% lower liver Cu concentrations in cattle (*Bos taurus*) supplemented with 0.58 mg Se/kg diet compared with 0.18 mg Se/kg in the control diet over a 7- to 8-month period. Koh and Judson (1987) evaluated the efficacies of oxidized Cu particles and Se bullets on the production responses in young cattle marginally deficient in both elements. The Se bullets exerted an antagonistic effect on the availability of Cu from the CuO particles, as indicated by reduced Cu concentrations in the whole blood, plasma, blood cells, and liver. Animal weight gains were significantly increased when both Cu and Se were supplemented, but not when provided individually. This has practical applications because Cu deficiency has been reported in areas where cattle and sheep (*Ovis aries*) are often supplemented with Se (H.F. Mayland, 1986, personal observation).

Sulfate S is another anion that interferes with Se absorption and metabolism. Halverson et al. (1962) reported that the addition of sodium sulfate to rat (*Rattus rattus*) diets reduced the toxicity of 10 mg Se as selenate per kilogram diet. However, adding the sulfate salt to diets containing selenite or seleniferous wheat was ineffective in reducing the toxicity. Thus, increasing the sulfate-S levels in diets that are marginally adequate in Se could be expected to increase the clinical incidence of white muscle disease (Hintz & Hogue, 1964). Pope et al. (1979) also found that blood-Se levels in sheep were inversely related to dietary S levels, but this effect could be countered by increasing the dietary intake of Se. Linseed oil meal has been found effective in reducing the toxicity of Se in rat diets (Palmer et al., 1980). The protective substances were cyanohydrin glycosides, which upon hydrolysis yield HCN. Conceivably, cyanide released from the glycosides might react with some of the Se compounds to form selenocyanates, which could be excreted by the animal. Therefore, excess levels of Se can be detoxified by feeding or injecting cyanide (Palmer & Olson, 1979), since Se and cyanide tend to counter the toxicity of the other.

# FECAL AND URINARY SELENIUM

Urine is the primary route of Se excretion by monogastric animals, regardless of whether the Se is given orally or injected. The main route of Se excretion in ruminants, though, is a function of the method of administration and the age of the animal (NAS-NRC, 1983). When Se is ingested by ruminants, most of it is excreted in feces. In contrast, Se that is injected either intravenously or subcutaneously into ruminants is excreted mostly in urine. Lambs, and presumably calves, that have not developed rumen function can excrete 66 to 75% of the orally ingested Se in the urine. Since most exogenous fecal Se is likely Se that has been reduced to an unavailable form such as elemental Se (Langlands et al., 1986), rumen organisms undoubtedly contributed to this age effect.

Nearly all of the Se excreted in the feces of ruminants is in an unavailable form and very little is available for uptake by plants. Peterson and Spedding (1963) showed that <0.3% of the Se taken up by three pasture species originated from the Se contained in sheep manure during a 75-d study. The manure was obtained from sheep dosed orally with isotopically labeled selenite.

Trimethylselenonium ion (TMSe<sup>+</sup>) is an important urinary Se metabolite. When added to nutrient solutions, TMSe<sup>+</sup> was readily absorbed and translocated to leaves and stems, but not to the grain of wheat (Olson et al., 1976a). However, large differences were observed in Se uptake by barley, wheat, and alfalfa when TMSe<sup>+</sup> was applied in a soil-pot study in the greenhouse. The authors noted that very little of the Se from TMSe<sup>+</sup> was absorbed by plants, and some of the absorbed TMSe<sup>+</sup> was even lost to the atmosphere through volatilization from the plant. Therefore, TMSe<sup>+</sup> excreted in animal urine likely contributes little biologically active Se to plants.

A portion of the Se added to soil as  $TMSe^+$  was also biologically volatilized, and this loss was increased by liming the soil at rates to increase the pH from 5.45 (initial) to values of pH 7.05 (Olson et al., 1976b). In addition, 30 to 50% of the Se added as  $TMSe^+$  to several different soils was sorbed to the soil particles during a 21-d period. The biologically inactive  $TMSe^+$  in urine plus the stimulation of plant growth by the N and S added in the form of urine may explain the lowered Se contents in grass growing on urine patches (Joblin & Pritchard, 1983).

## SELENIUM VOLATILITY

Volative Se compounds are naturally released into the atmosphere as a result of biological activity in aquatic (Chau et al., 1976) and terrestrial ecosystems (Abu-Erreish et al., 1968; Doran & Alexander, 1977). The pathways for Se volatilization in higher plants have been reviewed by Lewis (1976). The volatile compounds include dimethyl selenide (DMSe), dimethyl diselenide, dimethylselenone, methane selenol, and hydrogen selenide (Zieve & Peterson, 1981, 1984c).

Dimethyl selenide is the principle compound released by microbial and plant respiration (Lewis et al., 1966, 1974; Francis et al., 1974). Volatile Se was first collected from accumulator and nonaccumulator plants by Lewis et al. (1966, 1974), and the volatile Se respired by the accumulator A. racemosus was identified as DMSe (Lewis et al., 1974). Although DMSe was not found in the volatiles from a nonaccumulator Astragalus (Evans et al., 1968), failure to detect the DMSe in this latter case may possibly relate to detection levels or to differences in metabolism.

Seven cruciferous species gave off volatile Se during growth on nutrient media containing selenite or selenate. The rate of Se respiration varied from <0.05% of leaf-Se per hour to 0.40% per hour (Lewis, 1976). The DMSe was identified as the single volatile Se compound produced by fresh cabbage leaves.

Zieve and Peterson (1985) observed that <sup>75</sup>SeDMSe was also sorbed to soils. The amount sorbed was greater for wet soils than dry and was greater when the atmospheric concentration of DMSe was high than when it was low. Sorption of DMSe by individual soil constituents was greatest for organic matter and clays, less for Mn and Fe oxides, and least for acid-washed sands. Experiments with sterilized soils indicated that microorganisms were not significantly involved in the sorption process but may have been responsible for the conversion of DMSe to other Se forms after sorption occurred. They concluded that the soil was an important natural sink for atmospheric DMSe.

Under certain conditions, significant quantities of Se as DMSe are respired by animals as a volatile compound having a garlic-like odor (Martin, 1973; Sternberg et al., 1968). This compound is formed in animals by methylating selenite. The amount exhaled increases with increasing Se intake and is greater for orally administered selenite than selenate, selenomethionine, or seleniferous wheat (Martin, 1973). The formation of DMSe can be inhibited by As, Cd, and other heavy metals. This method of Se excretion is an important mechanism by which the animal removes Se from the body, especially when the diet is relatively high in Se.

Measurable concentrations of alkylselenides (principally DMSe) may now be detected in the atmosphere in some areas. Values of  $1 \text{ ng/m}^3$  have been noted by Zieve and Peterson (1984a, 1986) around swamps and landfills. Zieve and Peterson (1984a) have shown that DMSe in the atmosphere can be absorbed through the leaves of plants and transported to roots. This process occurred even when plants were grown in hydroponics with selenite-Se added to the nutrient culture.

# SELENIUM AS A FERTILIZER

Selenium fertilization is briefly discussed to provide information to interested agronomists and animal nutritionists.

The clay fraction and organic matter content in soil play a major role in the bioavailability of Se, especially selenite, which is readily sorbed to these soil constituents. Selenate is less readily sorbed in finer-textured soils or soils with higher organic matter concentrations. Consequently, selenate is usually absorbed from soils by plants more readily than is selenite (Carter et al., 1969). Greater absorption of selenate may also result from differences in root permeability or that selenite is reduced to elemental Se on the root surface. Although sulfate-S exerts little effect on selenite uptake, it competes with the absorption of selenate (Gissel-Nielsen, 1973; Westermann & Robbins, 1974).

The development of a safe and effective Se fertilizer must consider the chemical form, its solubility under the range in soil pH and redox potentials, and the rate and amount of Se uptake by the plant. The application of lime or companion fertilizer containing phosphate, sulfate, or N can also affect Se concentrations in the herbage (Gissel-Nielsen, 1973; Milchunas et al., 1983). These materials may contain small amounts of Se, or they may stimulate root growth and subsequent uptake of soil Se (Carter et al., 1972; Westermann & Robbins, 1974). Application of these nutrients often stimulate forage yields, thereby diluting the Se concentration in the forage (H.F. Mayland, 1975 unpublished data).

Fertilizers made from rock phosphates may contain as much as 200 mg Se/kg (Rader & Hill, 1935; Robbins & Carter, 1970; Senesi et al., 1979). These concentrations are generally much higher than those measured in fertilizers derived from rock carbonates, synthetic N fertilizers, and even potassium sulfate (Senesi et al., 1979). Thus, some fertilizer materials may contribute Se for use by higher plants.

Selenium-deficient soils are often S-deficient, especially for the production of legumes. The application of sulfate fertilizers can simultaneously stimulate plant growth and compete with soil selenate for absorption sites on the root and transport mechanisms in the plant. Increasing the concentrations of sulfate-S in the forage also may have detrimental effects on Se availability to the animal (Hintz & Hogue, 1964; Pope et al., 1979).

There is some risk associated with the application of selenates to acid and neutral soil because of their high bioavailability (Reuter, 1975). Experience has shown that the use of selenate fertilizer results in much higher levels (sometimes toxic to animals) of Se in the first cut of forages following fertilization (Tables 2-3 and 2-4). Selenium concentrations in the forage, fertilized with a selenate source, usually tend to decrease sharply with each subsequent harvest (Carter et al., 1969; Giessel-Nielsen & Bisbjerg, 1970; Ylaranta, 1983).

T	ble 2–4. Selenium uptake by barley straw and grain from soil-applied Se fertilizer, foli	ar-
	applied Se spray or Se-treated seed in 1980 and the residual response in 1981 on a F	in-
	nish soil having 6.1 pH, 2.5% organic carbon, and 41% clay (adapted from Ylaran	ta,
	1983).	

Sel	enium	Gr	ain	Sti	raw
Source	Rate	1980	1981	1980	1981
	g Se/ha		-mg Se/kg	dry matter—	
	N-P-F	K mix, field e	xperiment		
Control	0.	0.022	0.017	0.019	0.016
Na selenite	50. 200. 1050.	0.067 0.345 1.56	0.018 0.029 0.070	0.022 0.170 0.830	0.020 0.025 0.044
Na selenate	10. 50. 500. Foliar spray at 1	0.097 1.12 7.45 Feekes 4 sta	0.022 0.020 0.039 ge, field expe	0.054 0.813 6.38 riment	0.020 0.017 0.018
Control	0,	0.021	0.013	0.021	0.017
Na selenite	5. 50. 500.	0.041 0.183 1.11	0.021 0.018 0.023	0.052 0.126 0.923	0.018 0.018 0.024
Na selenate	5. 50. 500.	0.040 1.47 7.52	0.021 0.019 0.036	0.038 0.334 6.79	0.018 0.016 0.020
	Barley seed	treatment, p	ot experime	nt†	
Control Na selenite Na selenate	0. 15. 17.	0.020 0.042 0.305		0.017 0.018 0.200	

† Selenium rates are equivalent to the amount applied per hectare when treated seed is planted at the rate of 200 kg/ha.

The addition of selenite to acid or neutral soils results in immobilization of some Se. This process includes sorption of selenite, probably by hydrous sesquioxides, which reduces the solubility of Se to plants so that they can take up protective (for animal requirements), but not toxic levels of Se. The other major process resulting in the immobilization of Se is the reduction of selenite to elemental Se, selenide, or both. This reduced Se apparently is unavailable to plants in the short term and its oxidation to more available forms may be slow (Cary et al., 1967).

Foliar application of selenite to plants provides another means to increase Se in the diet of the animal. Gissel-Nielsen (1981) sprayed 3 to 5 g Se/ha as selenite with a wetting agent onto young barley plants at Feekes growth stage 4 to 6 and demonstrated that this was an effective way of getting Se into the grain. Selenium applied as  $Na_2SeO_3$  to alfalfa or ryegrass seed at the rate equivalent to 0, 50, 100, or 200 g Se/ha maintained plant tissue-Se levels at >0.1 mg/kg through three harvests (Gupta et al., 1984). Ylaranta (1983) reported that the Se concentration in barley grain could be increased by (i) blending selenite or selenate with N-P-K compounded fer-

tilizers and (ii) spraying plants at the midtillering stage (Feekes 4), but not by seed treatment (Table 2-4). On the basis of these investigations, one can conclude that selenite is a better source than selenate when the concentration in plants is to be raised to a sufficient but not excessive level for livestock nutrition.

The application of 10 g Se/ha as selenite to pastures has been suggested (Gissel-Nielsen, 1984). The Se is applied with a carrier fertilizer such as calcium ammonium phosphate. A method now legally used in New Zealand is the top-dressing of pastures with Se prills (Gissel-Nielsen, 1984). Korkman (1985) reported that only New Zealand and Finland had approved the addition of Se to fertilizers with which feedstuffs and food were produced. Contrary to the suggestions of Gissel-Nielsen, both countries use the selenate form at rates of about 10 g Se/ha. In New Zealand, 1% Se granulate is mixed with the granulated fertilizer in bulk blending equipment, whereas in Finland a 1% Se solution is sprayed into a granulation drum and Se is thus distributed into all fertilizer granules at an equivalent rate of 6 g Se/ha for silage and 16 g Se/ha for cereal. In New Zealand about  $0.5 \times 10^6$  ha and in Finland about  $2.3 \times 10^6$  ha would be treated annually, beginning in 1985.

# SELENIUM IN COAL AND MUNICIPAL REFUSE COMBUSTION

# **Redistribution of Selenium**

Natural weathering of the earth's crustal material results in an estimated  $1.0 \times 10^6$  kg and  $7.2 \times 10^6$  kg Se being mobilized annually and transported by rivers draining the land mass in the continental USA and world, respectively (Goldberg et al., 1971; Turekian, 1971). In addition, relatively small, but often detectable amounts of volatile Se compounds are released into the atmosphere as a result of biological activity in terrestrial and aquatic ecosystems (Peterson, 1980; Zieve & Peterson, 1984b). This small amount may be quite important on a global basis.

Human activities are significantly impacting the global Se cycle. Andren et al. (1975) concluded that 1.5 to 2.5 times as much Se is mobilized through coal burning as by natural weathering processes. Most coal consumed in the USA is burned at power plants where Se and other elements entering the boiler are partitioned between a bottom ash stream (slag) and a flue-gas stream containing suspended fly ash and the vapors of volatile elements or compounds. The flue-gas stream next passes through electrostatic precipitators, or other devices that efficiently remove (>95% wt./wt.) fly ash particles >1.5 to 2.0  $\mu$ m mass median aerodynamic diameter (MMAD). These scrubbers are less efficient in removing vapors and finer particles (Klein et al., 1975). Most of the bottom ash and the ash from the precipitators are transported to land fills as solid residues or flushed with water to ash ponds, where the Se may be leached from the ash and enter the aquatic environment. About 15% of the residue is recycled in building materials, roadways, concrete, etc. (Eisenberg et al., 1986). Small particles, <2  $\mu$ m MMAD, and

vapors are discharged as aerosols to the atmosphere, only to enter the terrestrial and aquatic environment by wet or dry deposition.

A Se mass balance study was conducted at the Allen Steam Plant in Memphis, TN (Andren et al., 1975). This plant was equipped with a high-efficiency electrostatic precipitator that allowed only 0.5% of the total incoming fly ash to be exhausted to the atmosphere. However, as much as 15 to 20% of the 0.1 to 0.5  $\mu$ m MMAD particles and a very large portion of the <0.1  $\mu$ m MMAD particles were lost as aerosols. Of the 2.2 mg Se/kg coal entering the boiler, 0.3% exited with the slag, 68% was with the precipitated fly ash particles, and the remaining 32% was associated with the vapor phase. The Se in the slag, fly ash, and vapor phases was identified as elemental Se. However, some of this Se must occur as or eventually become bioavailable, because fly ash is a good source of Se for higher plants (see section on Bioavailability of Fly Ash Selenium, below).

An estimated  $6 \times 10^{11}$  and  $28 \times 10^{11}$  kg of coal (having about-11% ash) are combusted annually in the USA and world, respectively (Andren et al., 1975). Perhaps coal consumption has been reduced or data were calculated differently, because Eisenberg et al. (1986) quoted a figure of 4.5  $\times 10^{10}$  kg of fly ash produced in the USA in 1980.

Upon combustion, about 40% of the ash occurs as bottom ash or slag, 60% as fly ash, and <1% as aerosols that escape to the atmosphere (Eisenberg et al., 1986). These proportions may differ for other coal-boiler plants, because of design and operating conditions. Selenium concentrations in these components at the Allen Steam Plant were 2.2, 0.08, 28, and 88 mg Se/kg for the bulk coal, slag, fly ash, and aerosol fractions, respectively.

# Selenium Enrichment Factors in Aerosols, Particulates, and Precipitation

The Se enrichment of a given fraction relative to crustal rocks provides a useful index to follow the preferential movement of Se (or other elements) in the environment. The enrichment factor (EF) for Se is calculated as follows:

EF = (Se/Al)aerosol/(Se/Al)crust

The term (Se/Al)aerosol is the ratio of the Se concentration to that of Al in the aersol and the (Se/Al)crust is the ratio of the elements in igneous rocks of the upper continental crust (Wedepohl, 1971). This approach assumes that Al is conserved in the combustion process and thus the EF for Al is 1.0. The EF values could also be calculated for the Se enrichment in the ash or other carrier. Similar EF values of 84 and 132 were calculated for Se in ash from 11 coal-fired power plants in the United Kingdom and 23 in the USA, respectively (Wadge et al., 1986; Furr et al., 1977).

Wadge et al. (1986) separated fly ash into size fractions ranging from 1.25 to 76  $\mu$ m MMAD. Most of the particles were <20  $\mu$ m MMAD. The Se concentrations in the bulk ash from the 11 plants ranged from 2 to 26 mg Se/kg ash with a mean of 5.4 mg Se/kg. These researchers reported that Se concentrations were inversely related to particle size and further that the

calculated EF of the 2  $\mu$ m MMAD particle was 2 times that of the 50- $\mu$ m particle. In another study conducted in the USA (Campbell et al., 1978), the EF for Se in the fine particles compared to larger particles was 9.9, illustrating that some variation occurs between coal plants or in sampling and analytical techniques.

High combustion temperatures volatilize some of the Se in the coal, which then leaves the furnace in the gaseous phase. Fly ash and gases begin to condense as they move through the particle scrubbers and ascend the stacks. Most of the larger particles and the Se associated with them are trapped by the scrubbers. However, the remaining Se probably condenses on the smaller particles that escape the stack and enter the atmosphere as aerosols. The degree of preferential enrichment may be a function of the individual power plant and current operating conditions. The amount of Se leaving the stack will be a function of the Se content of the coal, particle size, combustion temperature, efficiency of particle scrubbers, temperature of flue-gases, and exit velocity.

Another source of aerosols that can enter the atmosphere and contribute to global cycling of Se is the incineration of refuse. Nationally we generate  $16 \times 10^{10}$  kg of municipal refuse annually in the USA (Lemley, 1987). While incineration of wastes would reduce this mass, it would also be a source of aerosol and ash residues. In the United Kingdom, municipal refuse incineration consumed  $3 \times 10^9$  kg of material annually generating  $1 \times 10^9$ kg ash residue (Wadge et al., 1986). Unlike coal-fired power plants, more bottom ash is produced than fly ash. Refuse incinerators operate at lower temperatures (800-1100 °C) compared to coal furnaces that operate at nearly 1500 °C. Coal fly ash is composed of cenospheres of  $< 20 \ \mu m$  MMAD, whereas refuse fly ash consisted mostly of amorphous particles of  $< 50 \ \mu m$ MMAD.

Wadge et al. (1986) reported that refuse fly ash from an incinerator located in the United Kingdom contained 2.5 to 6.6 mg Se/kg with a mean value of 4.4. The Se in this refuse fly ash had an EF of 93 compared with an EF of 98 reported for refuse fly ash from an incinerator in the USA (Greenberg et al., 1978a). These EF values are similar to those given for coal-derived fly ash of 84 and 132 given above. Greenberg et al. (1987b) reported an EF of 4500 for Se in aerosols above a refuse incinerator in the USA, possibly reflecting the larger particle sizes and cooler incinerator temperatures when compared with data for coal furnaces (Andrean et al., 1975). Although the EF values of Se are similar for the two refuse incinerators, one should not assume that all municipal refuse is alike or their Se concentrations the same. Measurable differences may exist among Se concentrations in refuse from rural vs. industrial vs. metropolitan areas.

Chiou and Manuel (1986) collected atmospheric particulates at Rolla, MO, using a high-volume particle fractionating cascade impactor. This system gave five sized fractions with a range of 0.01 to >7.0  $\mu$ m MMAD particle. They measured 3.6  $\pm$  0.8  $\mu$ g Se/m<sup>3</sup> compared with a range of 2.1 to >7  $\mu$ g/m<sup>3</sup> measured elsewhere in the USA (Ondov et al., 1982; Pillay et al., 1971; Rahn & Lowenthal, 1985; Scheff et al., 1984). The EF for Se increased exponentially as particle size decreased. About 79% of the aerosol Se was associated with the  $< 2.0 \ \mu m$  MMAD particles.

The relative Se sorption to the particle is inversely related to particle size (Chiou & Manuel, 1986) and is likely a surface phenomena. A halving of the diameter of a sphere causes a doubling of the surface area for the same mass of particles. The particle distribution at Rolla, MO, may have been affected by the lead mines and smelters in the area. Most important is the fact that 70% of the particles collected by Chiou and Manuel (1986) were <2.0  $\mu$ m compared to the >2.0  $\mu$ m particles collected in other studies reported here.

The computed EF values for select elements in the bulk Missouri aerosol (particles of 0.01-7  $\mu$ m MMAD) were:

Tellurium	12 000
Selenium	2 200
Antimony	1 900
Sulfur	780
Arsenic	320
Zinc	110
Chlorine	56
Manganese	5.6
Calcium	1.7
Aluminum	1.0

Small EF values indicate little redistribution of the element among various materials, whereas the large EF values indicate considerable redistribution. The EF for the Se in the 0.01 to 1.1  $\mu$ m MMAD particles was about 50 000, which is much greater than the 2200 value calculated for the bulk aerosol.

Upon combustion, coal-derived Se is preferentially partitioned to the submicron-sized particles, which are easily transported over long distances. This might account for the high enrichment factors of 18 000 reported for Se aerosols at the South Pole (Zoller et al., 1974) and the similar EF of 22 000 reported at Memphis, TN (Andren et al., 1975). Although these values are greater than the value of 2200 calculated for the Se in aerosols captured in Missouri (Chiou & Manuel, 1986), the two previous aerosols likely contained much smaller particles than those from Missouri.

Using Se/S and Te/S ratios, Chiou and Manuel (1986) were able to show that coal combustion was the prime source of chalcogen elements (O, S, Se, and Te). They discounted the contribution of soil and volcanic ash to the Se found in the aerosols. Also, biological methylation of inorganic forms of Se in soil and sewage has been suggested as an important source of atmospheric Se (Reamer & Zoller, 1980). However, the amounts generated by these mechanisms are not well quantified and are considered to be much less than those from the combustion of coal and refuse (Chiou & Manuel, 1986).

Cutter & Church (1986) reported data on the concentration and oxidation state of Se in precipitation over coastal and midocean regions of the western Atlantic Ocean. The results indicate that fossil fuel combustion enriches Se in wet depositional fluxes to western Atlantic surface waters. They measured an average of 30 and 170 ng Se/L in rainwater samples collected over Bermuda and Lewes, DE, respectively. On occasion, values as high as 300 ng Se/L were encountered over the Lewes site. They calculated that the wet depositional flux at Lewes, DE, was about 15 ng Se/cm<sup>2</sup> per year (given as 190 pmol/cm<sup>2</sup> per year). That amount is equivalent to 1.5 g Se/ha annually, which is about one-tenth the amount of fertilizer Se that is applied annually in Se-deficient areas such as Finland and New Zealand. The wet depositional flux calculated by Cutter and Church (1986) falls between the values estimated by Ross (1985) for remote continental (75 pmol/cm<sup>2</sup> per year) and urban (840 pmol/cm<sup>2</sup> per year) wet deposition.

Selenium's oxidation state in precipitation may be a sensitive oxidationreduction (redox) tracer (Cutter & Church, 1986). The authors found that the range of selenite/selenate values was generally small [Se(IV)/Se(VI) =  $1.26 \pm 0.95$ ]. Two samples with the highest selenite/selenate ratios (15-20) came from storms traveling in northerly directions, placing the coal-fired Indian River Station power plant 30 km upwind of the rain sampler. They calculated an apparent redox intensity of the precipitation, but suggested that more data were needed to accurately determine the oxidation intensity of air masses.

Andren et al. (1975) reported that Se in the fly ash and aerosols retrieved from the Allen Steam Plant was all in the elemental form (Se°). Cutter and Church (1986) detected selenide plus elemental Se in only one sample accounting for 28% of the total Se. The remaining (72%) Se occurred as selenite and selenate. Fly ash has been shown to contain sufficient concentrations of bioavailable Se that it could serve as a supplemental nutrient source for plants and animals (Furr et al., 1975, 1976a, 1977, 1978).

Suzuki et al. (1981) and Cutter (1978) reported that selenite was the major Se species in rainwater samples collected in Japan and on the California coast. Robberecht and van Grieken (1980) reported variable quantities of both selenite and selenate in the rain and snow samples collected from an urban area in Belgium.

## **Bioavailability of Aerosol Selenium**

The chemical speciation of Se and the particle size of the carrier aerosol are important in assessing its bioavailability. Oral ingestion is the primary route of entry into the animal body for most trace elements, although inhalation of Pb may be an important route for body Pb loads. Aerosols may enter the body through the respiratory system and serve as another source of Se. While the larger 1 to 20  $\mu$ m MMAD particles are deposited primarily in the nasal and bronchial region of the respiratory system, the submicron particles are deposited predominantly in the lungs and have the greatest enrichment in many of the trace elements, including Se (Campbell et al., 1978).

Medinsky et al. (1981) modeled the distribution and retention of Se in rats after inhalation of elemental Se and selenious acid aerosols (0.6  $\mu$ m MMAD). They noted that the rate of absorption of these two Se forms into

the blood was different, but once absorbed, both chemical forms behaved identically. In a second study, Weissman et al. (1983) used beagle dogs (*Canis familiaris*) and calculated that the animals absorbed 52 and 73% of the Se from the Se metal and selenious acid aerosols, respectively. In this study the aerosols had 0.7 and 0.5  $\mu$ m MMAD, respectively. Selenium absorption from the Se<sup>0</sup> or H<sub>2</sub>SeO<sub>3</sub> forms was 73 and 96% when given by stomach pump and 10 and 50% when given in the feed, respectively. Selenium that was absorbed into the blood was translocated to various organs with a whole-body biological half-life of 34 d. Urine was the major route of excretion, accounting for 70 to 80% of the excreted Se.

Atmospheric deposition patterns, based on elemental concentrations in mosses, have been reported in Norway (Rambaek & Steinnes, 1980). More recently, Froslie et al. (1985) measured the trace element concentration in the livers of 6 to 8 month old lambs that had grazed on unfertilized mountain pastures at 11 different geographical areas in that country. They reported correlation coefficients for elemental concentrations in liver vs. atmospheric deposition of each element as follows:

These values strongly indicate a relationship between atmospheric deposition of several trace elements, including Se, and intake by grazing animals. The Se intake by the lambs may not only be due to the metal concentration sorbed by the plants from the atmospheric fallout but could be affected by ingestion of soil (Mayland et al., 1975), since surface soils showed a similar trace element contamination pattern. Froslie et al. (1985) concluded that the Se fallout from aerosol sources over southern and southwestern Norway played a beneficial role in reducing the incidences of Se-deficiency disorders in livestock.

Freshwater lakes or other waters may also receive Se by natural cycling processes or as discharges of wastes from irrigation drainage, like that at Kesterson in central California, or from coal-combustion or refuse incineration plants. Soluble Se forms (selenite, selenate) present in the water have a relatively high bioavailability and may be responsible for reduced reproduction (Gillespie & Baumann, 1986) and growth (Sorensen & Bauer, 1984; Sorensen, 1986) in fish. The Se passes along the food chain from algae to rotifer to larval fish to larger fish and to fish-eating birds and even humans (Bennett et al., 1986; Bertram & Brooks, 1986; Gissel-Nielsen & Gissel-Nielsen, 1978; Hodson et al., 1986; Chapter 8 of this publication). This aspect of Se cycling will require persistent monitoring and adoption of management strategies to reduce overall risks of Se toxicosis in plants and animals.

# **Bioavailability of Fly Ash Selenium**

Approximately 26 to  $33 \times 10^9$  kg fly ash were generated annually in the USA between 1975 and 1980 (Combs et al., 1980; Furr et al., 1976a). Some of this fly ash has been used as a soft aggregate in construction applications and as filter aids in processing municipal sewage. Because these applications consume only a small fraction of the total production, its possible use in agriculture has been investigated.

Fly ashes vary greatly, not only in particle size and elemental composition, but also in pH (Furr et al., 1977). Use of these ashes as soil amendments could affect the availability of Se and other elements, both in the ash and in the soil, and should be carefully evaluated. The pH of the ash, concentration of total soluble salts, and presence of elements toxic to plants are factors that affect growth and elemental concentration in the plant.

Several studies have shown that at least a portion of the Se in fly ash is available for uptake by plants grown naturally or experimentally on fly ash or on soils amended with fly ash (Combs et al., 1980; Furr et al., 1976a, 1978; Gutenmann et al., 1979; Stoewsand et al., 1978). Furr et al. (1976a) found that the amount of Se absorbed by the edible portion of several vegetables was proportional to the amount of coal-derived fly ash amendment added to the soil. In that study, the plants absorbed less Se during the second cropping than during the first. In another study (Gutenmann et al., 1979), the Se in fly ash was steadily released over five successive cuttings of alfalfa, birdsfoot trefoil (*Lotus corniculatus* L.), bromegrass, orchardgrass, and timothy.

Agricultural uses of refuse-incinerator fly ash may be more restricted than those for coal-derived fly ash. For example, the incinerator fly ash used in a study by Wadge and Hutton (1986) not only contained 4.3 mg Se/kg but also had high concentrations of Cd (450 mg Cd/kg) and Pb (8200 mg Pb/kg). These heavy metals can affect plant growth and have a negative impact on the nutritional value of the crops. Cabbage [*Brassica oleracea* (Capitata Group)] and barley were grown on soil containing from 0 to 40% (wt./wt.) of the incinerator fly ash. Plant growth was reduced by the fly ash, and the yield of the barley crop in the 40% amendment group was only about 30% of the control value. The first crop of cabbage and barley contained 0.51 and 1.08 mg Se/kg, respectively, when grown in media containing 20% ash (wt./wt.) compared with 0.01 and 0.07 mg Se/kg when grown in the control media not containing ash.

Corn (Zea mays L.) was grown in two successive years on soil amended with  $1 \times 10^6$  kg/ha (500 tons/acre) coal-derived fly ash (Combs et al., 1980). Use of the fly ash resulted in a sevenfold increase in total Se in corn grain produced during the two seasons after application of the fly ash. The Se in this corn had a bioavailability to chicks (Gallus gallus) of about 46% when compared with that of selenite.

Animals fed plants grown on fly ash or fly ash-amended soils have absorbed some of the Se as evidenced by increases in tissue Se concentrations (Furr et al., 1975, 1978; Stoewsand et al., 1978). Yellow sweet clover [Melilo-

tus officinallis (L.) Pall.], grown on fly ash containing 5.3 mg Se/kg, was formulated at 35% of a diet fed to hamsters (*Mesocricetus auratus*) and guinea pigs (*Cavia porcellus*) for 90 d (Furr et al., 1975). Sheep and goats (*Capra hircus*) were fed a pelleted diet containing seleniferous white sweet clover (*Melilotus alba* Medik.) harvested from a fly ash disposal area. The sweet clover contained 66 mg Se/kg, but as prepared contained only 15.5 mg Se/kg, which was fed for 173 d. In yet another study, wheat grain containing 5.7 mg Se/kg, was fed to Japanese quail (*Coturnix coturnix japonica*). The wheat had been grown on fly ash containing 21 mg Se/kg (Stoewsand et al., 1978). Feed consumption and weight gains in each of these studies were not adversely affected by the seleniferous diets. Although body tissues from animals fed the seleniferous diets contained elevated levels of Se, outward signs of selenosis were not evident.

# SELENIUM IN MUNICIPAL SEWAGE SLUDGE

Municipal sewage sludge represents a major portion of the national solidwaste disposal problem with an estimated  $4.5 \times 10^{10}$  kg produced in 1968 (Furr et al., 1976b). Furr et al. (1976c) measured the elemental concentrations in sludges collected from 16 U.S. cities and reported that the spectrum and concentration of heavy metals in the sludge was generally a function of the industry in the area. Selenium concentrations in the 16 sludges ranged from 1.7 to 8.7 mg Se/kg, with an average of 3.2 mg Se/kg.

The bioavailability of the Se (and other metals) becomes an important factor in the disposal of sewage sludges. A heat-treated, waste-activated sludge from the city of Milwaukee, WI, has been marketed commercially as Milorganite, a soil conditioner for application on lawns and gardens. The product contains 1.8 mg Se/kg. Furr et al. (1976b) grew a variety of garden crops in pots with soil containing 10% Milorganite by weight. At harvest, the edible portions of these crops had significantly greater concentrations of Se than for those plants grown on the control soil, but the increase may not have been biologically significant.

Waste from feed lots, poultry houses, and municipal sewage treatment plants have also been evaluated as a supplemental feed for livestock. The manures tested by Capar et al. (1978) contained 0.5 to 0.7 mg Se/kg, whereas the Denver municipal sludge contained 4.6 soluble and 7.2 mg/kg total Se. Feeding the Denver sludge as 0, 4, or 12% of diet to 340 kg steers provided diets containing 0.16, 0.35, or 0.72 mg Se/kg (Kienholz et al., 1979). The sludge provided very little metabolizable energy, and animal weight gains were less compared with control animals. There was a trend (P > 0.05) for increasing Se concentrations in liver; 1.3, 1.7, and 4.2 mg Se/kg (dry basis) for the three treatments, respectively. Selenium concentrations in dried blood were 1.3, 1.7, and 12 mg Se/kg, respectively (P < 0.05).

In another study (Boyer et al., 1981), steers were fed a basal or basal diet containing 12% Fort Collins anaerobically digested sewage sludge. The sludge contained 16 mg Se/kg but, after dilution with the rest of the ration,

	Se in tissues, mg/kg wet basis		
Tissue	Control	Sludge-amended	
Kidney	1.35	1.55	
Liver	0.42	1.05	
Muscle	0.23	0.30	
Spleen	0.36	0.39	
Brain	0.20	0.23	

Table 2-5. Selenium levels in tissues of cattle receiving a basal diet containing an estimated 0.16 mg Se/kg or a sludge-amended diet containing 1.7 mg Se/kg (adapted from Boyer et al., 1981).

provided 1.7 mg Se/kg of diet. Tissues obtained after slaughter of these animals showed small, but consistent, increases in Se concentration as a result of sludge supplementation, compared with tissues from animals on the basal diet (Table 2-5).

Although the energy-depleted sludges are not likely to be used in livestock rations because of reduced animal performance, these wastes could be used as soil conditioners in gardening, landscaping, and cropping situations where they provide organic matter and some plant nutrients. However, the Se concentrations found in municipal sludges tested to date are not threatening the quality of edible produce grown on sludge-amended soil.

# AGRONOMIC IMPACTS ON SELENIUM WEATHERING

# **Crop-Fallow Versus Seleniferous Cretaceous Materials**

Humans have also impacted the natural weathering rate of seleniferous materials by choice of cropping systems. This has occurred in many areas of the Great Plains that once were rangelands, but now are largely in dryland grain production. This type of agriculture has been possible because of the adoption of a crop-fallow system.

Fallowing in alternate years conserves extra water for use by the next year's grain crop. However, this cropping system, especially in years of above average precipitation, allows excess water to percolate beyond the root zone where it may contact the underlying Cretaceous geologic formations (Brown et al., 1983). The percolating water may then encounter impervious layers and move laterally to intersect the surface, forming a spring or seep. These seeps have a detrimental impact on the area because of their high salinity.

The Great Plains area is underlain with Cretaceous geological materials, including formations that are classified as being in the Montana or Colorado groups (Donovan et al., 1981; Miller et al., 1981). The seleniferous groundwater in the northern Great Plains is associated with the dark or black shales of the Colorado group where the unweathered shale may contain 0.25 to 10.3 mg Se/kg. Groundwater samples from these dark shales have correspondingly high Se values. The highest Se values were typically found in thin glacial till deposits and weathered shale overlying relatively

Component	Colorado	Montana	F ratio†
pH	7.6	7.9	7.1
-	me	/L	
Ca	275	120	37
Mg	866	71	41
Na	1317	670	12
К	17.7	6.1	NS
Fe	0.28	0.26	NS
SiO,	12.1	11.6	NS
HCÔ,	534	722	8
CI	141	177	NS
SO,	6041	1143	38
NO	57	6.1	32
Sr	4.6	2.0	26
TDS	9262	2928	32
Se	0.308	0.0276	35

Table 2-6. Water chemical-data for samples taken from the Cretaceous-aged Colorado or Montana Group formations in central Montana (Donavan et al., 1981).

 $\dagger F$  ratios for chemical constituents calculated in a one-way analysis of variance between data of the two groups. F = 6.75 or 11.1 at P = 0.01 or 0.001, respectively. NS = not significant, all F values <1.0.

unweathered shale in areas where saline-seep discharges occur. The hydrology of these *local* hydrologic systems, including both contributing and seep area, is described by Brown et al. (1983) and Donovan et al. (1981).

Water chemistries were determined on 81 samples collected from domestic, livestock, or research wells drilled into the cretaceous formations of central Montana (Donovan et al., 1981). The chemistry of each sample was closely related to the particular geological source (Table 2–6). Water samples from the Colorado group have higher proportions of Mg, SO<sub>4</sub>, NO<sub>3</sub>, and Se when compared with water obtained from the Montana formations. Selenium concentrations in these waters ranged from <2 to 1400  $\mu$ g Se/L.

# **Irrigation Versus Seleniferous Cretaceous Materials**

Humans have also impacted the Se cycle through the use of leaching fractions to remove excess salts from the root zone of irrigated agricultural crops. In the western part of the San Joaquin Valley, these leaching fractions have intercepted soluble Se initially derived from the Cretaceous Moreno shales (Tanji et al., 1986). Drains were installed to lower the groundwater table underlying the farmland. The drainage water contained soluble salts including Se, which were then drained to and concentrated in the evaporation ponds at the Kesterson Reservoir. The Se was now accessible to aquatic life and sufficiently available to alter the spectrum of bacterial genera present (Burton et al., 1987) and to cause chronic and acute toxicosis of the aquatic wildlife (see Chapter 8 of this publication).

# SUMMARY

Selenium is of biological importance because of (i) its essentiality in animal and possibly plant metabolism, (ii) its insufficiency for animal requirements in many areas, and (iii) its toxicity to animals and plants in other areas where it occurs in high concentrations in soil, water, plants, or in aerosols. Animals require 0.05 to 0.1 mg Se/kg in their diets to prevent Se deficiency, but suffer Se toxicosis when dietary levels exceed 5 to 15 mg Se/kg. If plants require Se, the critical level is <0.08 mg/kg.

Selenium occurs in several valency states controlled by the pH and Eh of the soil. The  $Se^{6+}$ , and to a lesser extent  $Se^{4+}$ , salts are water soluble and are the predominant forms absorbed by plants. Animals easily absorb selenate and to a lesser extent selenite sources, but seem to best utilize the amino and proteinaceous forms found in plants.

The earth's crustal materials generally contain <0.1 mg Se/kg. Higher concentrations are found in Cretaceous shales. The Se-accumulator plants growing on seleniferous soils may contain hundreds or even thousands of mg Se/kg. However, the nonaccumulator grasses and forbs seldom accumulate >50 mg Se/kg and more often contain <5 mg Se/kg.

Anthropogenic activities impact the amount of Se entering our nation's lakes, rivers, and the atmosphere. Combustion of coal and incineration of municipal waste exhaust Se into the environment. In addition, crop-fallow and irrigation practices that allow leaching waters to pass through seleniferous strata, prior to intersecting with surface flow, augment the Se levels encountered by various segments of plant and animal life.

An improved understanding of Se in the agroecosystem will allow us to constructively manage this element in areas where it is now deficient for animal nutrition and other areas where it exists at levels leading to toxicosis in animals and plants.

# ACKNOWLEDGMENTS

The authors thank Drs. W.H. Allaway, E.E. Cary, A.C. Chang, O.E. Olson, M. Coburn Williams, and the anonymous reviewers for their helpful comments. The senior author especially thanks Dr. I.S. Palmer for allowing access to his extensive file of Se literature.

### REFERENCES

Abuereish, G.M., and J.N. Lahham. 1987. Selenium in soils and plants of the Jordan Valley. J. Arid Environ. 12:1-7.

Abu-Erreish, G.M., E.I. Whitehead, and O.E. Olson. 1968. Evolution of volatile selenium from soils. Soil Sci. 106:415-420.

Adriano, D.C. 1986. Trace elements in the terrestrial environment. Springer-Verlag, New York.

- Allaway, W.H. 1968. Agronomic controls over the environmental cycling of trace elements. Adv. Agron. 20:235-274.
- Allaway, W.H., E.E. Cary, and C.F. Ehlig. 1967. The cycling of low levels of selenium in soils, plants and animals. p. 273-296. In O.H. Muth (ed.) Symposium: Selenium in biomedicine. AVI Publishing Co., Westport, CT.
- Anderson, J.W., and A.R. Scarf. 1983. Selenium and plant metabolism. p. 241-275. In D.A. Robb and W.S. Pierpoint (ed.) Metals and micronutrients: Uptake and utilization by plants. Academic Press, New York.

- Anderson, M.S., H.W. Lakin, K.C. Beeson, F.F. Smith, and E. Thacker. 1961. Selenium in agriculture. USDA Agric. Handb. 200. U.S. Gov. Print. Office, Washington, DC.
- Andren, A.W., D.J. Klein, and Y. Talmi. 1975. Selenium in coal-fired steam plant emissions. Environ. Sci. Technol. 9:856-858.
- Bennett, W.N., A.S. Brooks, and M.E. Boraas. 1986. Selenium uptake and transfer in an aquatic food chain and its effects on fathead minnow larvae. Arch. Environ. Contam. Toxicol. 15:513-517.
- Bertram, P.E., and A.S. Brooks. 1986. Kinetics of accumulation of selenium from food and water by fathead minnows. Water Res. 20:877-884.
- Boyer, K.W., J.W. Jones, D. Linscott, S.K. Wright, W. Stroube, and W. Cunningham. 1981. Trace element levels in tissues from cattle fed a sewage sludge-amended diet. J. Toxicol. Environ. Health 8:281-295.
- Brown, P.L., A.D. Halvorson, F.H. Siddoway, H.F. Mayland, and M.R. Miller. 1983. Salineseep diagnosis, control, and reclamation. Conserv. Res. Rep. U.S. Dep. Agric. 30:1-22.
- Brown, T.A., and A. Shrift. 1982. Selenium: Toxicity and tolerance in higher plants. Biol. Rev. 57:59-84.
- Broyer, T.C., C.M. Johnson, and R.P. Huston. 1972a. Selenium and nutrition of Astrogalus. I. Effects of selenite or selenate supply on growth and selenium content. Plant Soil 36:635-649.
- Broyer, T.C., C.M. Johnson, and R.P. Huston. 1972b. Selenium and nutrition of Astragalus. II. Ionic sorption interactions among selenium, phosphate, and the macro- and micronutrient cations. Plant Soil 36:651-669.
- Broyer, T.C., D.C. Lee, and C.J. Asher. 1966. Selenium nutrition of green plants. Effect of selenite supply on growth and selenium content of alfalfa and subterranean clover. Plant Physiol. 41:1425-1428.
- Burton, G.A., Jr., T.H. Giddings, P. DeBrine, and R. Fall. 1987. High incidence of seleniteresistant bacteria from a site polluted with selenium. Appl. Environ. Microbiol. 53:185-188.
- Campbell, J.A., J.C. Laul, K.K. Nielson, and R.D. Smith. 1978. Separation and chemical characterization of finely-sized fly-ash particles. Anal. Chem. 50:1032-1040.
- Capar, S.G., J.T. Tanner, M.H. Friedman, and K.W. Boyer. 1978. Multielement analysis of animal feed, animal waste, and sewage sludge. Environ. Sci. Technol. 12:785-790.
- Carter, D.L., M.J. Brown, and C.W. Robbins. 1969. Selenium concentrations in alfalfa from several sources applied to a low selenium, alkaline soil. Soil Sci. Soc. Am. Proc. 33:715-718.
- Carter, D.L., C.W. Robbins, and M.J. Brown. 1972. Effect of phosphorus fertilization on the selenium concentration in alfalfa (*Medicago sativa*). Soil Sci. Soc. Am. Proc. 36:624-628.
- Cary, E.E., G.W. Wieczorek, and W.H. Allaway. 1967. Reactions of selenite-selenium added to soils that produce low selenium forages. Soil Sci. Soc. Am. Proc. 31:21-26.
- Chau, Y.K., P.T.S. Wong, B.A. Silverberg, P.L. Luxon, and G.A. Bengert. 1976. Methylation of selenium in the aquatic environment. Science (Washington, DC) 192:1130-1131.
- Chiou, K.Y., and O.K. Manuel. 1986. Tellurium and selenium in aerosols. Environ. Sci. Technol. 20:987-991.
- Combs, G.F., Jr., S.A. Barrows, and F.N. Swader. 1980. Biologic availability of selenium in corn grain produced on soil amended with fly ash. J. Agric. Food Chem. 28:406-409.
- Combs, G.F., Jr., and S.B. Combs. 1986. The role of selenium in nutrition. Academic Press, New York.
- Cutter, G.A. 1978. Species determination of selenium in natural waters. Anal. Chim. Acta 98:59-66.
- Cutter, G.A., and T.M. Church. 1986. Selenium in western Atlantic precipitation. Nature (London) 322:720-722.
- Davies, E.B., and J.H. Watkinson. 1966. Uptake of native and applied selenium by pasture species. N.Z. J. Agric. Res. 9:317-327.
- Donovan, J., J.L. Sonderegger, and M.R. Miller. 1981. Investigations of soluble salt loads, controlling mineralogy, and factors affecting the rates and amounts of leached salts. Montana Water Resour. Res. Center Rep. 120. Montana State Univ., Bozeman, MT.
- Doran, J.W., and M. Alexander. 1977. Microbial formation of volatile selenium compounds in soil. Soil Sci. Soc. Am. J. 41:70-73.
- Ehlig, C.F., W.H. Allaway, E.E. Cary, and J. Kubota. 1968. Differences among plant species in selenium accumulation from soils low in available selenium. Agron. J. 60:43-47.
- Eisenberg, S.H., M.E. Tittlebaum, H.C. Eaton, and M.M. Soroczak. 1986. Chemical characteristics of selected flyash leachates. J. Environ. Sci. Health Part A 21:383-402.

- Ellrich, J., A. Hirner, and H. Stark. 1985. Distribution of trace elements in crude oils from southern Germany, Chem. Geol. 48:313-323.
- Evans, C.S., C. Asher, and C.M. Johnson. 1968. Isolation of dimethyl diselenide and other volatile selenium compounds from Astragalus racemosus (Pursh.). Aust. J. Biol. Sci. 21:13-20.
- Food and Nutrition Board. 1980. Recommended dietary allowances. NAS-NRC, Washington. DC, p. 162-164.
- Francis, A.J., J.M. Duxbury, and M. Alexander. 1974. Evolution of dimethylselenide from soils. Appl. Microbiol. 28:248-250.
- Frank, A., B. Pehrson, and L.R. Petersson. 1986. Concentration of some important elements in the liver of young cattle supplemented with selenite enriched feed. J. Vet. Med. Assoc. 33:422-425.
- Froslie, A., G. Norheim, J.P. Rambaek, and E. Steinnes. 1985. Heavy metals in lamb liver: Contribution from atmospheric fallout. Bull. Environ. Contam. Toxicol. 34:175-182.
- Furr, A.K., W.C. Kelly, C.A. Bache, W.H. Gutenmann, and D.J. Lisk. 1976a. Multielement uptake by vegetables and millet grown in pots on fly ash amended soil. J. Agric. Food Chem. 24:885-888.
- Furr, A.K., W.C. Kelly, C.A. Bache, W. Gutenmann, and D.J. Lisk. 1976b. Multielement absorption by crops grown in pots on municipal sludge-amended soil. J. Agric. Food Chem. 24:889-892.
- Furr, A.K., A.W. Lawrence, S.S.C. Tong, M.C. Grandelle, R.A. Hofstader, C.A. Bache, W.H. Gutenmann, and D.J. Lisk. 1976c. Multielement and chlorinated hydrocarbon analysis of municipal sewage sludges of American cities. Environ. Sci. Technol. 10:683-687.
- Furr, A.K., T.F. Parkinson, C.L. Heffron, Jr., J.T. Reid, W.M. Haschek, W.H. Gutenmann, C.A. Bache, and J.E. St. John. 1978. Elemental content of tissues and excreta of lambs, goats and kids fed white sweet clover growing on fly ash. J. Agric. Food Chem. 26:847-851.
- Furr, A.K., T.F. Parkinson, R.A. Hinrichs, D.R. Van Campen, C.A. Bache, W.H. Guten-mann, L.E. St. John, and I.S. Pakkala. 1977. National survey of elements and radioactivity in fly ashes. Absorption of elements by cabbage grown in fly ash-soil mixtures. Environ. Sci. Technol. 11:1194-1201.
- Furr, A.K., G.S. Stoewsand, C.A. Bache, W.A. Gutenmann, and D.J. Lisk. 1975. Multielement residues in tissues of guinea pigs fed sweet clover grown on fly ash. Arch. Environ. Health 30:244-248.
- Gillespie, R.B., and P.C. Baumann. 1986. Effects of high tissue concentrations of selenium on reproduction by bluegills. Trans. Am. Fish. Soc. 115:208-213.
- Gissel-Nielsen, G. 1973. Uptake and distribution of added selenite and selenate by barley and red clover as influenced by sulfur. J. Sci. Food Agric. 24:649-655.
- Gissel-Nielsen, G., 1984. Improvement of selenium status of pasture crops. Biol. Trace Ele-ment Res. 6:281-288.
- Gissel-Nielsen, G. 1981. Foliar application of selenite to barley plants low in selenium. Commun. Soil Sci, Plant Anal. 12:631-642.
- Gissel-Nielsen, G., and B. Bisbjerg. 1970. The uptake of applied selenium by agricultural plants. 2. The utilization of various selenium compounds. Plant Soil 32:382-396.
- Gissel-Nielsen, G., U.C. Gupta, M. Lemand, and T. Westermarck. 1984. Selenium in soils and plants and its importance in livestock and human nutrition. Adv. Agron. 37:397-461.
- Gissel-Nielsen, M., and G. Gissel-Nielsen. 1978. Sensitivity of trout to chronic and acute exposure to selenium. Agric. Environ. 4:85-91.
- Goldberg, E.D., W.S. Broeker, M.G. Gross, and K.K. Turekian. 1971. Marine Chemistry. In Radioactivity in the marine environment. NAS-NRC, Washington, DC.
- Greenberg, R.R., G.E. Gordon, W.H. Zoller, R.B. Jacko, D.W. Neuendorf, and K.J. Yost. 1978a. Composition of particles emitted from the Nicosia municipal incinerator. Environ. Sci. Technol. 12:1329-1332.
- Greenberg, R.R., W.H. Zoller, and G.E. Gordon. 1978b. Composition and size distributions of particles released in refuse incineration. Environ. Sci. Technol. 12:566-573.
- Gupta, U.C., K.A. Winter, and H.T. Kunelius. 1984. Effect of selenium treated seed on the
- Selenium concentration of forages. Can J. Plant Sci. 64:432 (abstract).
   Gutenmann, W.H., I.S. Pakkala, D.J. Churey, W.C. Kelly, and D.J. Lisk. 1979. Arsenic, boron, molybdenum, and selenium in successive cuttings of forage crops field grown on fly ash amended soil. J. Agric. Food Chem. 27:1393-1395.
- Halverson, A.W., P.L. Guss, and O.E. Olson. 1962. Effect of sulfur salts on selenium poisoning in the rat. J. Nutr. 77:459-464.

- Hamilton, J.W., and O.A. Beath. 1963. Uptake of available selenium by certain range plants. J. Range Manage. 16:261-264.
- Hamilton, J.W., and O.A. Beath. 1964. Amount and chemical form of selenium in vegetable plants. Agric. Food Chem. 12:371-374.
- Hintz, H.F., and D.E. Hogue, 1964. Effect of selenium, sulfur and sulfur amino acids on nutritional muscular dystrophy in the lamb. J. Nutr. 82:495-498.
- Hodson, P.V., J.W. Hilton, and S.J. Slinger. 1986. Accumulation of waterborne selenium by rainbow trout (Salmo gairdneri), eggs, fry and juveniles. Fish Physiol. Biochem. 1:187-196.
- Howard, J.H., 111. 1977. Geochemistry of selenium: Formation of ferroselite and selenium behavior in the vicinity of oxidizing sulfide and uranium deposits. Geochim. Cosmochim. Acta 41:1665-1678.
- Ihnat, M. 1989. Occurence and distribution of selenium. CRC Press, Boca Raton, FL.
- Jensen, N.L. 1985. Selenium, p. 1-7. In USDI Bureau of Mines. Bull, 675. U.S. Gov. Print. Office, Washington, DC.
- Joblin, K.N., and M.W. Pritchard. 1983. Urinary effect of variations in the selenium and sulphur contents of ryegrass from pasture. Plant Soil 70:69-76.
- Kabata-Pendias, A., and H. Pendias. 1984. Trace elements in soils and plants. CRC Press, Boca Raton, FL.
- Kienholz, E.W., G.M. Ward., D.E. Johnson, J. Baxter, G. Braude, and G. Stern. 1979. Metropolitan sewage sludge fed to feedlot steers. J. Anim. Sci. 48:735-741.
- Klein, D.H., A.W. Andren, J.A. Carter, J.F. Emery, C. Feldman, W. Fulkerson, W.S. Lyon, J.C. Ogle, Y. Talmi, R.I. van Hook, and N. Bolton. 1975. Pathways of thirty-seven trace elements through coal-fired power plant. Environ. Sci. Technol. 9:973-979.
- Koh, T.-S., and G.J. Judson. 1987. Copper and selenium deficiency in cattle: An evaluation of methods of oral therapy and an observation of a copper-selenium interaction. Vet. Res. Commun. 11:133-148.
- Koller, L.D., and J.H. Exon. 1986. The two faces of selenium—deficiency and toxicity—are similar in animals and man. Can. J. Vet. Res. 50:297-306.
- Korkman, J. 1985. Selenium in fertilizers, Bull. 28. Selenium-Tellurium Dev. Assoc., Darien, CT.
- Kubota, J., W.H. Allaway, D.L. Carter, E.E. Cary, and V.A. Lazar. 1967. Selenium in crops in the United States in relation to selenium-responsive diseases of animals. J. Agric. Food Chem. 15:448-453.
- Lakin, H.W. 1948. Selenium occurrence in certain soils of the United States, with a discussion of related topics: Seventh report. USDA Tech. Bull. 953. U.S. Gov. Print. Office, Washington, DC.
- Lakin, H.W. 1972. Selenium accumulation in soils and its absorption by plants and animals. Geol. Soc. Am. Bull. 83:181-189.
- Lakin, H.W., and H.G. Byers. 1948. Selenium occurrence in certain soils in the United States, with a discussion of related topics: Seventh report. USDA Tech. Bull. 960. U.S. Gov. Print. Office, Washington, DC.
- Lakin, H.W., and D.F. Davidson. 1967. The relation of the geochemistry of selenium to its occurrence in soils. p. 27-56. In O.H. Muth (ed.) Selenium in biomedicine. Van Nostrand Reinhold Co., New York.
- Langlands, J.P., J.E. Bowles, G.E. Donald, and A.J. Smith. 1986. Selenium excretion in sheep. Aust. J. Agric. Res. 37:201-209.
- Lemley, B. 1987. Garbage. Parade Magazine. 14 June, p. 20.
- Levander, O.A. 1986. Selenium. p. 209-280. In W. Mertz (ed.) Trace elements in human and animal nutrition. 5th ed. Vol. 2. Academic Press, New York.
- Lewis, B.G. 1976. Selenium in biological systems, and pathways for its volatilization in higher plants. p. 389-409. In J.O. Nriagu (ed.) Environmental biogeochemistry. Ann Arbor Science, Ann Arbor, M1.
- Lewis, B.G., C.M. Johnson, and T.C. Broyer. 1974. Volatile selenium in higher plants: The production of dimethyl selenide in cabbage leaves by enzymatic cleavage of Se-methyl selenomethionine selenonium salt. Plant Soil 40:107-118.
- Lewis, B.G., C.M. Johnson, and C.C. Delwiche. 1966. Release of volatile selenium compounds. by plants. Collection procedures and preliminary observations. J. Agric. Food Chem. 14:638-640.
- Martin, J.L. 1973. Selenium compounds in nature and medicine. B. Selenium assimilation in animals. p. 663-691. In D.L. Klayman and W.H.H. Gunther (ed.) Organic selenium compounds: Their chemistry and biology. Wiley Interscience, New York.

- Massey, D.L., and P.J. Martin. 1975. Selenium: Soil-plant-animal relationship. Alberta Agric. Agdex 531:1-4.
- Mayland, H.F., A. Florence, R.C. Rosenau, V.A. Lazar, and H.A. Turner. 1975. Soil ingestion by cattle on semiarid range as reflected by titanium analysis of feces. J. Range Manage. 28:448-452.
- Medinsky, M.A., R.G. Cuddiby, W.C. Griffith, and R.O. McClellan. 1981. A simulation model describing the metabolism of inhaled and ingested selenium compounds. Toxicol. Appl. Pharmacol. 59:54-63.
- Milchunas, D.G., W.K. Lauenroth, and J.L. Dodd. 1983. The interaction of atmospheric and soil sulphur on the sulphur and selenium concentration of range plants. Plant Soil 72:117-125.
- Miller, M.R., P.L. Brown, J.J. Donovan, R.N. Bergatino, J.L. Sonderegger, and F.A. Schmidt. 1981. Saline seep development and control in the North American great plains hydrogeological aspects. Agric. Water Manage. 4:115-141.
- Moxon, A.L., and M. Rhian. 1943. Selenium poisoning. Physiol. Rev. 23:305-337.
- National Academy of Science-National Research Council. 1974. Geochemistry and the environment: Vol. I. The relation of selected trace elements to health and disease. NAS-NRC, Washington, DC.
- National Academy of Science-National Research Council. 1976. Selenium. Assembly of Life Sciences. NAS-NRC, Washington, DC.
- National Academy of Science-National Research Council, 1983. Selenium in nutrition. Rev. ed. Board on Agric. NAS-NRC, Washington, DC.
- Olson, O.E., E.E. Cary, and W.H. Altaway. 1976a. Absorption of trimethylselenonium by plants. Agron. J. 68:805-809.
- Olson, O.E., E.E. Cary, and W.H. Allaway. 1976b. Fixation and volatilization by soils of selenium from trimethylselenonium. Agron. J. 68:839-843.
- Olson, O.E., E.J. Novacek, E.I. Whitehead, and I.S. Palmer. 1970. Investigations on selenium in wheat. Phytochemistry 9:1181-1188.
- Ondov, J.M., W.H. Zoller, and G.E. Gordon. 1982. Trace element emissions on aerosols from motor vehicles. Environ. Sci. Technol. 16:318-328.
- Palmer, I.S., and O.E. Olson. 1979. Partial prevention by cyanide of selenium poisoning in rats. Biochem. Biophys. Res. Commun. 90:1379-1386.
- Palmer, I.S., O.E. Olson, A.W. Halverson, R. Miller, and C. Smith. 1980. Isolation of factors in linseed oil meal protective against chronic selenosis in rats. J. Nutr. 110:145-150.
- Peterson, P.J. 1980. Turnover of arsenic and selenium in the environment. p. 1-25. In Reactions and interactions in biogeochemical cycles. Univ. of Warwick, Conventry, England.
- Peterson, P.J., L.M. Benson, and R. Zieve. 1981. Effect of heavy metal pollution on plants. p. 279-342. In N.W. Lepp (ed.) Effect of trace metals on plant function. Applied Science Publishers, New York.
- Peterson, P.J., and G. Butler. 1962. The uptake and assimilation of selenite by higher plants. Aust. J. Biol. Sci. 15:126-146.
- Peterson, P.J., and D.J. Spedding. 1963. The excretion by sheep of Se-75 incorporated into red clover: The chemical nature of the excreted selenium and its uptake by three plant species. N.Z. J. Agric. Res. 6:13-22.
- Pillay, K.K.S., C.C. Thomas, and J.A. Sondel. 1971. Activation-analysis of airborne as a possible indicator of atmospheric sulfur pollutant. Environ. Sci. Technol. 5:74-77.
- Pope, A.L., R.J. Moir, M. Somers, E.J. Underwood, and C.L. White. 1979. The effect of sulphur on 75-Se absorption and retention in sheep. J. Nutr. 109:1448-1455.
- Rader, L.F., and W.L. Hill. 1935. Occurrence of selenium in natural phosphates, superphosphates and phosphoric acid. J. Agric. Res. 51:1071-1083.
- Rahn, K.A., and D.H. Lowenthal. 1985. Pollution aerosol in the northeast-, northeastern-, and midwestern-contributions. Science (Washington, DC) 228:275-284.
- Rambaek, J.P., and E. Steinnes. 1980. Atmospheric deposition of heavy metals studied by analysis of moss samples using neutron activation analysis and atomic absorption spectrometry. p. 175-180. In Nuclear methods in environmental and energy research (Conf-800433). U.S. Dep. of Energy, Washington, DC.
- Reamer, D.C., and W.H. Zoller. 1980. Selenium biomethylation products from soil and sewage sludge. Science (Washington, DC) 208:500-502.
- Reuter, D.J. 1975. Selenium in soils and plant: A review in relation to selenium deficiency in South Australia. Agricultural Record 2:44-50.

- Robberecht, H., and R. van Grieken. 1980. Selenium content and speciation in environmental waters determined by x-ray fluorescence spectroscopy. Trace Subst. Environ. Health 14:362-366.
- Robbins, C.W, and D.L. Carter. 1970. Selenium concentration in P fertilizer materials and associated uptake by plants. Soil Sci. Soc. Am. Proc. 34:506-509.
- Rosenfeld, I., and O.A. Beath. 1964. Selenium. Geobotany, biochemistry, toxicity and nutrition. Academic Press, New York.
- Ross, H.B. 1985. An atmospheric selenium budget for the region 30-degrees-N to 90-degrees-N. Tellus 37:78-90.
- Rumble, M.A. 1985. Quality of water for livestock in man-made impoundments in the Northern High Plains. J. Range Manage. 38:74-77.
- Scheff, P.A., R.A. Wadden, and R.J. Allen. 1984. Development and validation of a chemicalelement mass balance for Chicago. Environ. Sci. Technol. 18:923-931.
- Senesi, N., M. Polemio, and L. Lorusso. 1979. Content and distribution of arsenic, bismuth, lithium and selenium in mineral and synthetic fertilizers and their contribution to soil. Commun. Soil Sci. Plant Anal. 10:1109-1126.
- Sharma, S., and R. Singh. 1983. Selenium in soil, plant, and animal systems. CRC Crit. Rev. Environ. Control 13:23-50.
- Shrift, A. 1958. Biological activities of selenium compounds. Bot. Rev. 24:550-583.
- Shrift, A. 1969. Aspects of selenium metabolism in higher plants. Ann. Rev. Plant Physiol. 20:475-494.
- Shrift, A. 1973. Selenium compounds in nature and medicine. E. Metabolism of selenium by plants and microorganisms. p. 763-814. In D.L. Klayman and W.H.H. Gunther (ed.) Organic selenium compounds: Their chemistry and biology. Wiley Interscience, New York.
- Singh, M., and N. Singh. 1978. Selenium toxicity and its detoxication by phosphorus. Soil Sci. 126:255-262.
- Smith, G.S., and J.H. Watkinson. 1984. Selenium toxicity in perennial ryegrass and white clover. New Phytol. 97:557-564.
- Smith, R.A., R.B. Alexander, and M.G. Wolman. 1987. Water-quality trends in the Nation's river. Science (Washington, DC) 235:1607-1615.
- Soltanpour, P.N., and S.M. Workman. 1980. Use of NH<sub>4</sub>HCO<sub>3</sub>-DTPA soil test to assess availability and toxicity of selenium to alfalfa plants. Commun. Soil Sci. Plant Anal. 11:1147-1156.
- Sorensen, E.M.B. 1986. The effects of selenium on freshwater teleosts. p. 59-116. In E. Hodgson (ed.) Reviews in environmental toxicology 2. Elsevier, New York.
- Sorensen, E.M.B., and T.L. Bauer. 1984. A correlation between selenium accumulation in sunfish and changes in condition factor and organ weight. Environ. Pollut. Ser. A 34:357-366.
- Sternberg, J., J. Brodeur, A. Imbach, and A. Mercier. 1968. Metabolic studies of seleniated compounds. III. Lung excretion of Se-75 and liver function. Int. J. Appl. Radiat. Isot. 19:669-684.
- Stoewsand, G.S., W.H. Gutenmann, and D.J. Lisk. 1978. Wheat grown on fly ash: High selenium uptake and response when fed to Japanese quail. J. Agric. Food Chem. 26:757-759.
- Suzuki, Y., Y. Sugimura, and Y. Miyake. 1981. The content of selenium and its chemical form in rain water and aerosol in Tokyo. J. Meterol. Soc. Jpn. 59:405-409.
- Swaine, D.J. 1955. The trace-element content of soils. Tech. Commun. 48. Commonw. Bur. of Soil Sci., Harpenden, England.
- Tanji, K., A. Lauchli, and J. Meyer. 1986. Selenium in the San Joaquin Valley. Environment 28:6-11, 34-39.
- Trelease, S.F., and H.M. Trelease. 1938. Selenium as a stimulating and possibly essential element for indicator plants. Am. J. Bot. 25:372-380.
- Trelease, S.F., and H.M. Trelease. 1939. Physiological differentiation in Astrogatus with reference to selenium. Am. J. Bot. 26:530-535.
- Turekian, K.K. 1971. Rivers, tributaries and estuaries. p. 9-73. In D.W. Hood (ed.) Impingement of man on the oceans. Wiley Interscience, New York.
- U.S. Environmental Protection Agency. 1977. National interim primary drinking water regulations. USEPA Rep. 57019-76-003. U.S. Gov. Print. Office, Washington, DC.
- van Dorst, S.H., and P.J. Peterson. 1984. Selenium speciation in the soil solution and its relevance to plant uptake. J. Sci. Food Agric. 35:601-605.
- Volcani, R., A. Bondi, Y. Lewin, and Ch. Neumark. 1957. Maintenance of milk cows in a selenium affected area. Refu. Vet. 13:188-192.

- Wadge, A., and M. Hutton. 1986. The uptake of cadmium, lead and selenium by barley and cabbage grown on soils amended with refuse incinerator fly ash. Plant Soil 96:407-412.
- Wadge, A., M. Hutton, and P.J. Peterson. 1986. The concentrations and particle size relationships of selected trace elements in fly ashes from U.K. coal-fired power plants and a refuse incinerator. Sci. Total Environ. 54:13-27.
- Wedepohl, K.H. 1971. Geochemistry. Holt, Rinehart, and Winston, New York.
- Weissman, S.H., R.G. Cuddihy, and M.A. Medinsky. 1983. Absorption, distribution, and retention of inhaled selenious acid and selenium metal aerosols in beagle dogs. Toxicol. Appl. Pharmacol. 67:331-337.
- Wen, F.N., C. Li, and C.-S. Chen. 1988. Ubiquity of selenium containing tRNA in plants. Plant Sci. 57:185-193.
- Westermann, D.T., and C.W. Robbins. 1974. Effect of SO<sub>4</sub> fertilization on Se concentration of alfalfa (*Medicago sativa L.*). Agron. J. 66:207-208.
- Williams, C., and I. Thornton. 1972. The effect of soil additives on the uptake of molybdenum and selenium from soils from different environments. Plant Soil 36:395-406.
- Ylaranta, T. 1983. Effect of applied scienite and scienate on the scienium content of barley (Hordeum vulgare), Ann. Agric. Fenn. 22:164-174.
- Zieve, R., and P.J. Peterson. 1981. Factors influencing the volatilization of selenium from soil. Sci. Total Environ. 19:277-284.
- Zieve, R., and P.J. Peterson. 1984a. The accumulation and assimilation of dimethylselenide by four plant species. Planta 160:180-184.
- Zieve, R. and P.J. Peterson. 1984b. Volatilization of selenium from plants and soils. Sci. Total Environ. 32:197-202.
- Zieve, R., and P.J. Peterson. 1984c. Dimethylselenide—an important component of the biogeochemical cycling of selenium. Trace Subst. Environ. Health 18:262-267.
- Zieve, R., and P.J. Peterson. 1985. Sorption of dimethylselenide by soils. Soil Biol. Biochem. 17:105-107.
- Zieve, R., and P.J. Peterson. 1986. An assessment of the atmosphere as a source of plant selenium: Application of stable and radioactive selenium isotopes. Toxicol. Environ. Chem. 11:313-318.
- Zoller, W.H., E.S. Gladney, and R.A. Duce. 1974. Atmospheric concentrations and sources of trace metals at the South Pole. Science (Washington, DC) 183:198-200.