Rhizomania; *Beet necrotic yellow vein virus* Storage rot; *Athelia*-like sp., *Botrytis cinerea*, and *Penicillium* spp. I. A. Eujayl and C. A. Strausbaugh, USDA-ARS NWISRL, 3793 N. 3600 E., Kimberly, ID 83341

## Kimberly sugar beet germplasm evaluated for rhizomania and storage rot resistance in Idaho, 2020.

Eight sugar beet (Beta vulgaris L.) lines from the USDA-ARS Kimberly sugar beet program and five check cultivars were screened for resistance to Beet necrotic yellow vein virus (BNYVV), the causal agent of rhizomania, and to storage rot. The rhizomania evaluation was conducted at the USDA-ARS North Farm in Kimberly. ID which has Portneuf silt loam soil and had been in barley in 2019. In the spring the field was plowed and fertilized (110 lb N and 120 lb  $P_2O_5/A$ ) and roller harrowed on 27 Mar 20. The germplasm was planted (density of 51,840 seeds/A) on 20 Apr. The plots were one row 10-ft long with 22-in. between-row spacing and arranged in a randomized complete block design with 6 replicates. The crop was managed according to standard cultural practices for southern Idaho. The trial relied on endemic field inoculum for rhizomania and storage rot development. The plots were rated for rhizomania foliar symptom (percentage of plants with yellow, stunted, upright leaves) development on 24 Aug. The plants were mechanically topped and hand harvested on 13-14 Oct. At harvest, ten roots per plot were rated for rhizomania symptom development using a scale of 0 to 9 (0 = healthy and 9 = dead; Plant Disease 93:632-638), with disease index (DI) treated as a continuous variable. At harvest, eight roots per plot were also placed in a mesh-onion bag and kept in an indoor commercial storage facility (temperature set point 34°F) in Paul, ID on 15 Oct. On 22 Feb 21, after 131 days in storage, the roots were evaluated for the percentage of root surface area covered by fungal growth or rot. Except for root ratings, data were analyzed in SAS (Ver. 9.4) using the general linear model (Proc GLM) procedure, and Fisher's protected least significant difference ( $\alpha = 0.05$ ) was used for mean comparisons. The root ratings were rank transformed prior to analysis with mixed linear models (Proc MIXED), but the non-transformed means have been presented in the table.

Rhizomania symptom development was uniform and other disease problems were not evident in the plot area. The BNYVV susceptible check plots (Check 1 and Red beet) had 100% foliar symptoms and high root disease ratings. Resistant check 3 had 1% foliar symptoms and a low root rating which indicates that resistance based on two genes is holding up. Single gene resistance (Checks 2 and 4) had foliar ratings ranging from 8 to 13% indicating single gene resistance is not completely effective, but the root ratings were still good. Based on the root ratings, all entries had some level of BNYVV resistance since they were all better than the susceptible checks. However, only entry 9 had a root rating similar to the resistant checks and the yellowish leaves (100% foliar rating) were likely an indication of nutrient deficiency and not RZ susceptibility. Four entries (1, 4, 9, and 10) had resistance to fungal rots in storage. Entry 1 has also had no foliar symptoms and a good storage rating in two previous evaluations. Entry 4 should be evaluated again next year since it had no foliar symptoms and a good storage rating. Some entries may serve as a starting point for identifying additional sources of resistance to BNYVV and storage rots.

Entry <sup>z</sup>	Description	Root rot in storage (%) <sup>y</sup>	RZ foliar rating (% susceptible plants)	RZ root rating <sup>x</sup>
Check 3	BTSSALCHK3 ( $R_z I R_z I R_z 2 R_z 2$ ) = $R_z I + R_z 2$ resistant check	5 e	1 fg	17 ј
9	KDH19-17, doubled haploid breeding line	11 с-е	100 a	19 h-j
Check 2	BTSSALCHK2 ( $R_z 2R_z 2$ ) = $R_z 2$ resistant check	18 bc	8 ef	21 hi
Check 4	BTSSALCHK4 ( $R_z I R_z I$ ) = $R_z I$ resistant check	11 с-е	13 e	22 h
1	KEMS12-FP17, inbred line selected from PI672570	4 e	0 g	24 e-g
7	ELRIL-125, RIL from Red Beet X Sugar Beet	21 ab	0 g	26 ef
5	KEMS12/KPS19 breeding population - F <sub>3</sub> families	22 ab	26 d	26 ef
2	KEMS12/KPS24 breeding population $-F_4$ families	16 b-d	37 c	27 de
4	KEMS12/KPS03 breeding population - F <sub>3</sub> families	8 de	0 g	29 b-d
3	KEMS12/KPS25 breeding population $- F_4$ families	29 a	0 g	30 bc
10	KDH39-33/KDH13 (PI663862) breeding population – F <sub>4</sub> families	9 de	86 b	32 b
Check 1	BTSSALCHK1 ( $rzrz$ ) = susceptible sugar beet check	21 ab	100 a	39 a
Red beet	Early Wonder $(rzrz)$ = susceptible red beet check	ND	100 a	48 a
$P > F^{w}$		< 0.0001	< 0.0001	< 0.0001
LSD		9	8	Trans

<sup>z</sup> All lines were *Beta vulgaris* subsp. *vulgaris*. Five commercial cultivars were included as checks.

<sup>y</sup>Root rot in storage = the percent of root surface area covered by fungal growth or rot. Fungal growth was dominated by an *Athelia*-like basidiomycete (Mycologia 104:70-78), *Penicillium expansum* Link, and *Penicillium cellarum* C. A. Strausb. & Dugan. Trace levels of *Botrytis cinerea* Pers. were also present. ND = no data.

<sup>x</sup> Ten roots per plot were evaluated for rhizomania symptoms using a scale of 0-9 (0 = healthy and 9 = dead; Plant Disease 92:581-587). Root rating = a disease severity index value for each plot established using the following formula:

[((A)0+(B)1+(C)2+(D)3+(E)4+(F)5+(G)6+(H)7+(I)8+(J)9)/90]100, where A-J are the number of plants in categories 0-9, respectively. Trans = the root ratings were rank transformed prior to analysis with mixed linear models, but the non-transformed means have been presented in the table.

<sup>w</sup>P > F was the probability associated with the F value. LSD = Fisher's protected least significant difference value ( $\alpha = 0.05$ ). Within a column, means followed by the same letter did not differ significantly based on Fisher's protected LSD.