Phosphorus Fertilization of Sugarbeets

D. T. WESTERMAN, G. E. LEGGETT, and J. N. CARTER

Received for publication June 1, 1976

Sugarbeets (Beta vulgaris L.) have been fertilized with phosphorus (P) in the U.S.A. for the past 30 to 40 years. During this period, numerous studies on P fertilization rates and application methods have been conducted. Much of this information has been summarized (12, 14, 15, 4). Recent soil test data from England and the U.S.A. suggest that the available P levels in many soils are sufficient for maximum root and sucrose production without P fertilization. Detrimental effects of P fertilization at excessive soil P levels have been reported (6, 5) but not all data support these conclusions (12). Increasing fertilizer costs have also made it essential that growers have adequate guidelines upon which to base fertilizer applications. Information has been limited in Idaho for establishing an adequate soil test P level for optimum sugarbeet production. With this as background, we conducted two field experiments evaluating 1) the P fertilizer requirements of sugarbeets at different soil test P levels, and 2) the effects of P fertilization on soils already containing adequate available P levels.

Methods and Materials

Field experiments were conducted in 1972 and 1973 on the Portneuf silt loam (Xerollic Calciorthid) soil. This soil has a weakly cemented hardpan beginning at the 16- to 18-in depth which restricts downward root growth but not water movement. The residual soil P levels (Table 1) in 1972 resulted from a P fertilization experiment conducted on dry beans in 1971, whereas those in 1973 were established by rototilling into the soil various P fertilizer rates in the fall of 1972 after the harvest of a spring wheat crop. The bean straw residue from 1971 and the wheat straw residue from 1972 were plowed down in the spring, prior to the 1972 and 1973 experiments, respectively. The spring applications of N (34-0-0) and P (CSP, 0-45-0) fertilizers were incorporated into the seedbed before planting. The amount of N fertilizer applied was determined by the method outlined by Carter, et al. (3). Soil samples were taken from each residual P treatment in 9-in increments to the 18-in depth before the spring fertilizer application. The samples were air-dried and the soil test P level (STPL) was measured by the
Table 1.—Effect of P level on root and sucrose yields, impurity indexes, and P uptake by sugarbeets.

<table>
<thead>
<tr>
<th>Experiment</th>
<th>1972</th>
<th>1973</th>
<th>1974</th>
</tr>
</thead>
<tbody>
<tr>
<td>Residual soil test P level (ppm)</td>
<td>6</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>P fertilizer rate (lb/A)</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Root yield</td>
<td>18.5</td>
<td>18.5</td>
<td>18.5</td>
</tr>
<tr>
<td>Sucrose</td>
<td>43.5</td>
<td>43.5</td>
<td>43.5</td>
</tr>
<tr>
<td>Impurity index</td>
<td>1.7</td>
<td>1.7</td>
<td>1.7</td>
</tr>
<tr>
<td>P uptake</td>
<td>123.5</td>
<td>123.5</td>
<td>123.5</td>
</tr>
</tbody>
</table>

Results and Discussion

Soil test phosphorus levels

Significant (P ≤ 0.01) root yield responses to P fertilization were observed at the lowest STPL each year (Table 1). Root yields were maximum when 60 lb P/A were applied at the 3.8 ppm STPL in 1972, whereas only 20 lb P/A were necessary at 6.6 ppm in 1973. In both experiments, root yields were not significantly increased by P fertilization when the STPL was greater than 10 ppm. However, yields increased significantly when the STPL increased from 10.8 to 30.6 in 1973 without P fertilization. The STPL in the 9- to 18-in soil layer was 2 to 3 ppm P in both experiments (data not shown). A higher STPL in this lower soil layer would probably decrease the STPL at which sugarbeets would respond to P fertilization but we did not confirm this in these experiments. Our results verify an earlier reported experiment on this same soil series (9). Similar data for other calcareous soils have been reported in Washington (7, 6), England (9), and in the western U.S. and Canada (11). However, some yield re-

NaHCO₃ method (11). Initial soil and beet petiole NO₃-N levels were determined by the NO₃-N specific-ion electrode method (10).

Sugarbeets were planted in 24-in rows in early April and hand-thinned to a within-row plant spacing of 8- to 10-in in late May. The experimental areas were furrow-irrigated when tensiometers installed in the row at the 18-in depth indicated that approximately 55% of the available soil moisture had been used.

Twenty petioles from the most recently matured blades were taken randomly from each plot near July 1, 15, and 30, and again near September 1. The petioles were separated from the blades and cut into 0.5-in sections. In 1973, whole plants (tops and storage roots) were also randomly sampled at thinning for dry-weight determinations. All plant samples were dried at 65°C and ground to pass a 40-mesh sieve. The petioles were extracted with 2% acetic acid for soluble PO₄-P (19, 1). Total-P concentrations were determined after wet ashing plant samples with a mixture of HClO₄ and HNO₃. Both P forms were determined by the vanadomolybdate method (8).

Root yields were estimated by hand-harvesting four 10-ft row sections from each plot in the 1972 experiment and by machine-harvesting four 35-ft row sections from each plot in the 1973 experiment. Samples of sugarbeet tops and crowns from two uniform 10-ft row sections were harvested from each plot in both experiments immediately prior to root harvest for determining total-P uptake. The impurity index (2) and sucrose content were determined on two randomly selected root samples (approximately 25 lb each) from each plot by a sugar company. The root pulp (collected during the sucrose analysis) and the top and crown samples were dried at 65°C, ground to pass a 40-mesh sieve, and analyzed for total P. The crown's P content has been added to that of the tops for this discussion.
petiole PO₄-P concentration decreases below 750 ppm between thinning and harvest (19, 15). Our data showed that a petiole PO₄-P concentration of 750 ppm early in the growing season may still limit yields if the decrease in petiole PO₄-P concentration during the season is not considered. In general, we found that petiole PO₄-P concentrations should be greater than 1200 ppm in early July and 700 ppm in late August to insure maximum yields (Figure 1). We also found agreement was good between the petiole PO₄-P concentration and the STPL (Figure 2). At 10 ppm STPL, the petiole PO₄-P concentrations were approximately 1200 ppm in early July and 700 ppm in late August.

![Figure 2](image2.png)

Figure 2.—Relationships between different STPLs and petiole PO₄-P concentrations.

The relationships between the total P concentration in the blades and root yields are shown in Figure 3. Using the same root yield criteria as in Table 1 to separate significant root yield differences, P concentrations above 0.24% in early July and 0.21% in late August were adequate. Very little information is available relating the total P concentration of blades to root yields, but petiole P concentrations from 0.25 to 0.27% have been reported necessary for maximum root yields (7, 6). Critical P concentrations in young seedling blades have been given as 0.42% total P (16); however, photosynthesis has been shown to decrease at P levels below 0.60% (17).

Total P uptake ranged from a low of 18.5 to over 30 lb P/A (Table 1). Approximately 50% of this remains on the field in the tops and crowns after the sugarbeet roots are harvested. Total P uptake was lower in 1972 compared with 1973 at equivalent sugarbeet root yields. The P uptake per ton of sugarbeet roots varied from 0.7 to 1.2 lb P/A under deficient and excessive levels of P, respectively.
Pectole NO₃-N levels in late August averaged 3900 and 1900 ppm in the 1972 and 1973 experiments, respectively. This indicates that the N level did not limit root yields in either experiment but may have been slightly high for maximum sucrose production (3). We found that all the other nutritive elements in the blades were at sufficient concentrations.

**Early plant growth**

We observed a visual top growth response to P fertilization early in the growing season in each experiment, but many of these differences had disappeared by early August. Early plant weight was related to P fertilization, particularly at the 6.6 ppm STPL (Figure 4). Phosphorus fertilization had very little effect on plant weights above a 10.8 ppm STPL. Plants were still larger at higher STPLs, but these did not significantly increase final root yields. Other researchers have reported that root yield improvements from P fertilization occurred early in the growing season (15, 5). This early growth effect was confirmed in 1973 by relating plant weights at thinning to final root yields (Figure 4). Increasing the early plant size by P fertilization or with higher STPLs increased the final root yields; however, no significant root or sucrose yield increases occurred when the plants were larger than 11 g/25 plants at thinning.

**Summary**

Phosphorus fertilizer applications did not significantly increase sugarbeet root yields when the NaHCO₃-soil test P level was greater than 10 ppm in the 0- to 9-in plow layer. Neither P fertilization nor soil test P level influenced the sucrose concentration or impurity indexes. Thus, sucrose yields increased linearly with root yields.

**Figure 3**—Relationship between total P in the most recently matured blade and root yields.

**Figure 4**—Relationship between seedling dry weights at thinning (31 May 1973) and root yields (A) and P levels (B) in the 1973 experiment.

Sugarbeets did not respond to P fertilization if the pectoles from the most recently matured blades contained more than 1200 ppm PO₄-P in early July and 700 ppm PO₄-P in late August. Under similar conditions, the blades contained 0.24 and 0.21% total P. The whole plant dry weights at thinning were related to final root yields, indicating that the yield potential resulting from P fertilization is established early in the growing season.

**Acknowledgments**

The authors thank the Amalgamated Sugar Company for the sucrose and impurity index analysis of the root samples, and S. E. Crothers and F. L. Roberts for their assistance in both the field and laboratory parts of this study.

**Literature Cited**


